

Modeling & Simulation (Computational Immunology)

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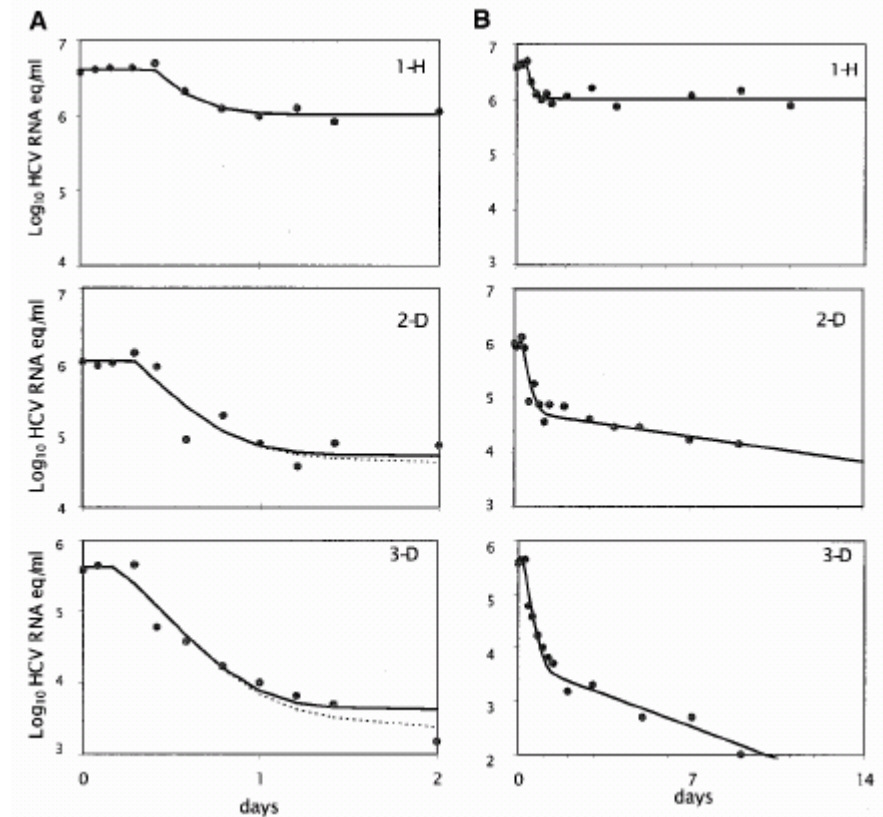
Hepatitis C Viral Dynamics and Interferon- α Therapy

Modeling 23 patients during 14 days of therapy (daily doses)

Hepatitis C Viral Dynamics in Vivo and the Antiviral Efficacy of Interferon- α Therapy

Avidan U. Neumann,*† Nancy P. Lam,*† Harel Dahari,
David R. Gretch, Thelma E. Wiley, Thomas J. Layden,
Alan S. Perelson

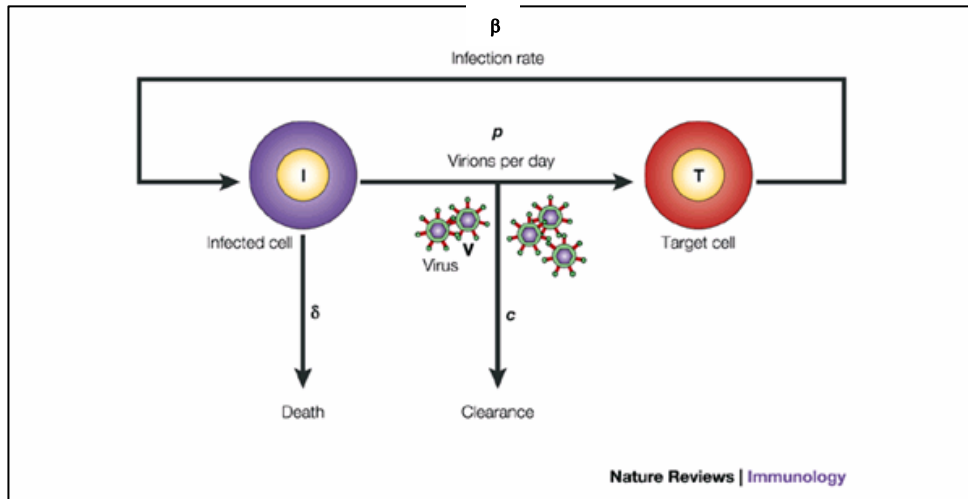
SCIENCE VOL 282 2 OCTOBER 1998



Short delay followed by biphasic decline in viral load

Model of Hepatitis C Viral Dynamics

Includes virus along with target (T) and infected (I) cells



Target Cells
$$dT/dt = s - dT \quad \boxed{\quad ? \quad} \quad (1)$$

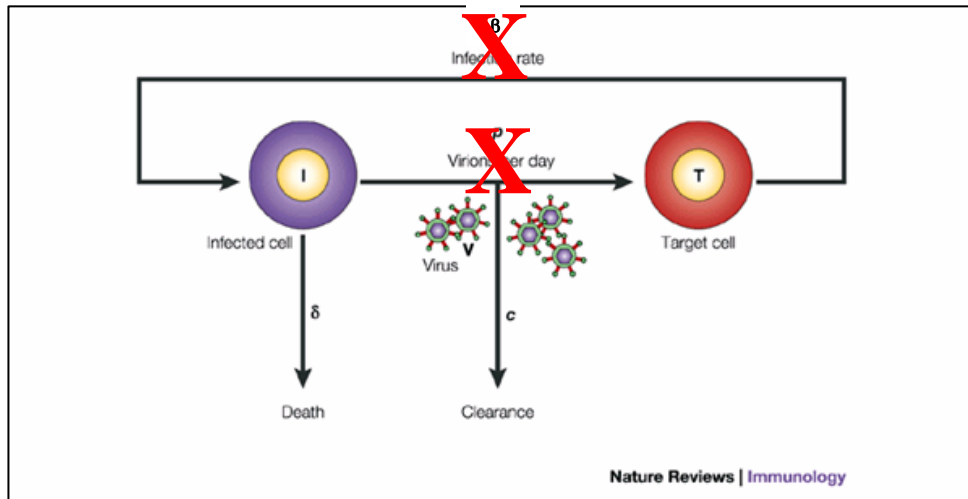
Infected Cells
$$dI/dt = \boxed{\quad ? \quad} - \delta I \quad (2)$$

Virus (HCV RNA)
$$dV/dt = pI - cV \quad (3)$$

Before therapy, virus load is approximately constant

Model of Interferon- α Therapy

Includes virus along with target (T) and infected (I) cells



Target Cells $dT/dt = s - dT - \beta VT$ (1)

Infected Cells $dI/dt = \beta VT - \delta I$ (2)

Virus (HCV RNA) $dV/dt = pI - cV$ (3)

Therapy can reduce the rate of infection, or production of virions

Hepatitis C Viral Dynamics and Interferon- α Therapy

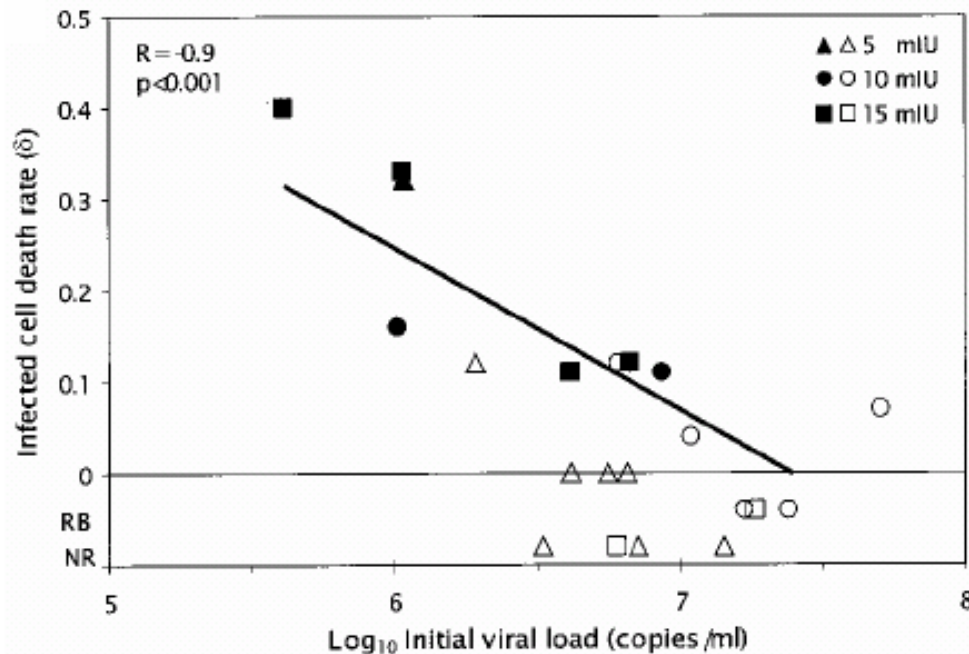
Modeling 23 patients during 14 days of therapy (daily doses)

Regimen	Patient	Initial VL (10^6 copies per milliliter)	Delay (hours)	Virion clearance (c)		Efficacy (ϵ)		Infected cell death (δ)		Production (10^9 copies per day)
				(1/day)	\pm error	Percent	\pm error	(1/day)	\pm error	
1	A	5.6	8	5.9	1.1	79	4.0%	0	0.01	495
1	B	1.9	8	6.4	1.8	75	7.0%	0.12	0.02	290
1	C	14.2	NR		NR		NR		NR	NR
1	D	7.1	NR		NR		NR		NR	NR
1	E	1.1	11	7.0	0.6	86	0.1%	0.32	0.04	125
1	F	6.5	7	5.0	0.8	89	8.0%	0	0.01	601
1	G	3.3	NR		NR		NR		NR	NR
1	H	4.1	10	6.9	0.2	75	1.0%	0	0.01	498
1: Mean	\pm SD	5.5 \pm 4.1	9 \pm 1.5	6.2 \pm 0.8		81 \pm 8%		0.09 \pm 0.14		402 \pm 191
2	A	6.1	7	3.6	0.2	86	0.5%	0.12	0.01	410
2	B	16.7	9	6.0	0.3	98	0.4%		RB	1409
2	C	8.6	8	6.8	0.8	96	1.0%	0.11	0.03	1089
2	D	1.0	7	5.6	0.5	95	1.0%	0.16	0.04	92
2	E	59.0	10	11.2	0.6	99.7	0.01%	0.07	0.02	12191
2	F	10.9	7	4.4	0.1	96	0.9%	0.04	0.01	965
2	G	23.8	7	4.8	0.1	92	0.8%		RB	1780
2	H	2.7	9	7.9	1.0	99.3	0.2%		ND	324
2: Mean	\pm SD	16.1 \pm 18.9	8 \pm 1	6.3 \pm 2.4		95 \pm 4%		0.1 \pm 0.05		2282 \pm 4045
3	A	6.7	8	3.7	0.3	99.7	0.4%	0.12	0.04	405
3	B	4.1	11	9.5	3.7	91	2.0%	0.11	0.03	761
3	C	5.8	13	5.7	0.7	98	0.5%		ND	523
3	D	0.4	5	6.0	0.8	99.0	0.2%	0.4	0.05	42
3	E	18.3	7	6.0	0.9	97.5	1.6%		RB	2136
3	F	1.1	14	5.8	0.6	90	0.3%	0.33	0.03	112
3	G	6.0	NR		NR		NR		NR	NR
3: Mean	\pm SD	6.0 \pm 5.9	9.5 \pm 3.5	6.1 \pm 1.9		96 \pm 4%		0.24 \pm 0.15		663 \pm 769
All: Mean	\pm SD	9.4 \pm 12.4	8.7 \pm 2.3	6.2 \pm 1.8		—		0.14 \pm 0.13		1276 \pm 498

Average virion production rate of 1.3×10^{12} virions per day

Hepatitis C Viral Dynamics and Interferon- α Therapy

Modeling 23 patients during 14 days of therapy (daily doses)



Suggests immune control has important role in lowering viral load

Patients with undetectable HCV after 3 months of therapy (filled symbols) had significantly faster cell death rates

Immune System Adapts to Pathogenic Challenge

Secondary responses are quantitatively and qualitatively different

**Faster kinetics,
greater magnitude**

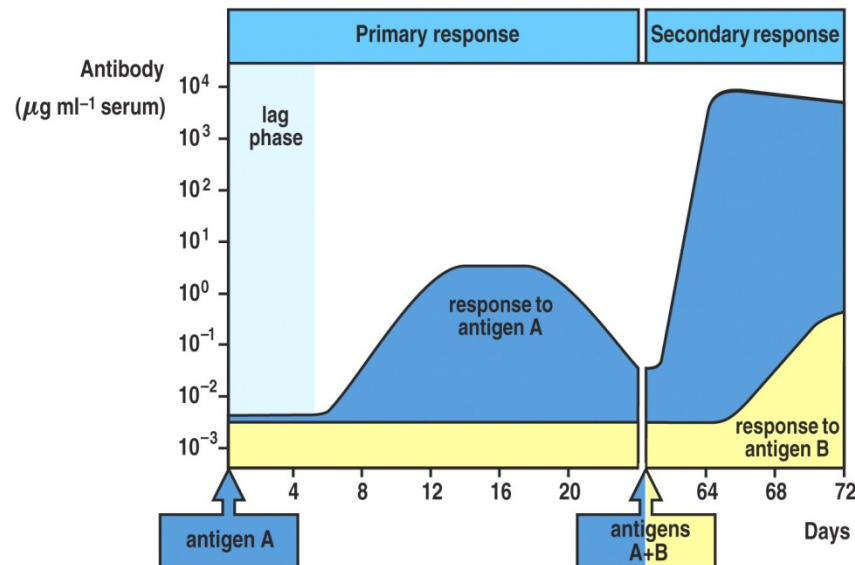


Figure 1-20 Immunobiology, 6/e. (© Garland Science 2005)

Increased affinity

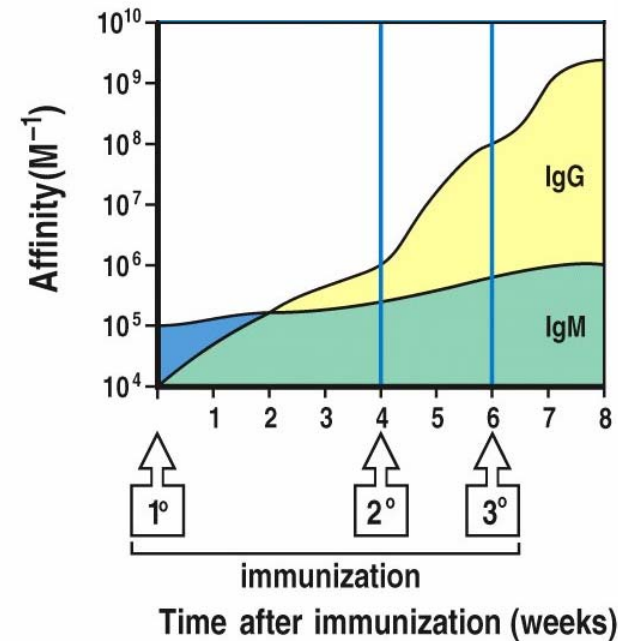
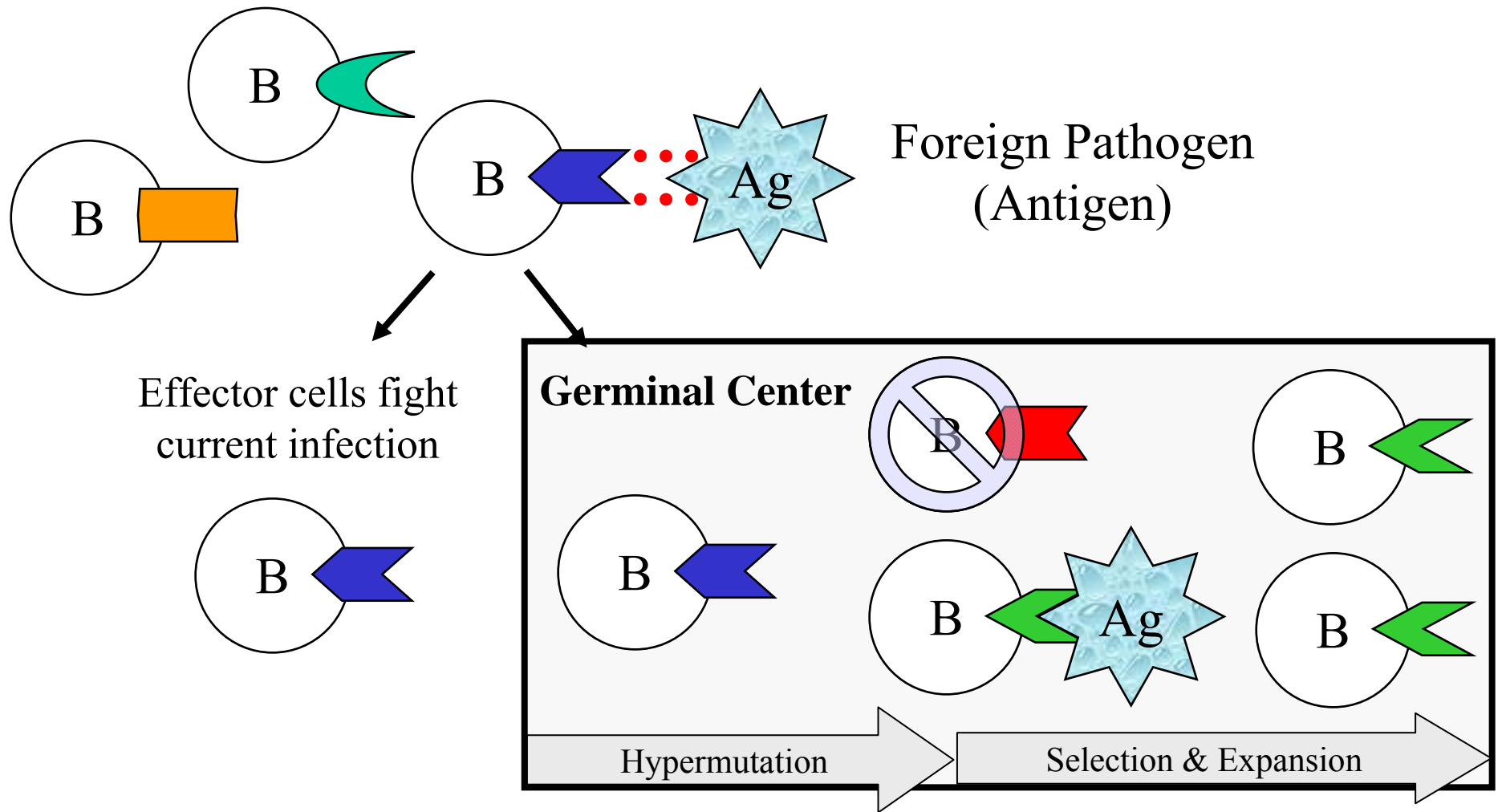


Figure 10-31 Immunobiology, 6/e. (© Garland Science 2005)

Affinity Maturation is Fundamental to Adaptive Immunity

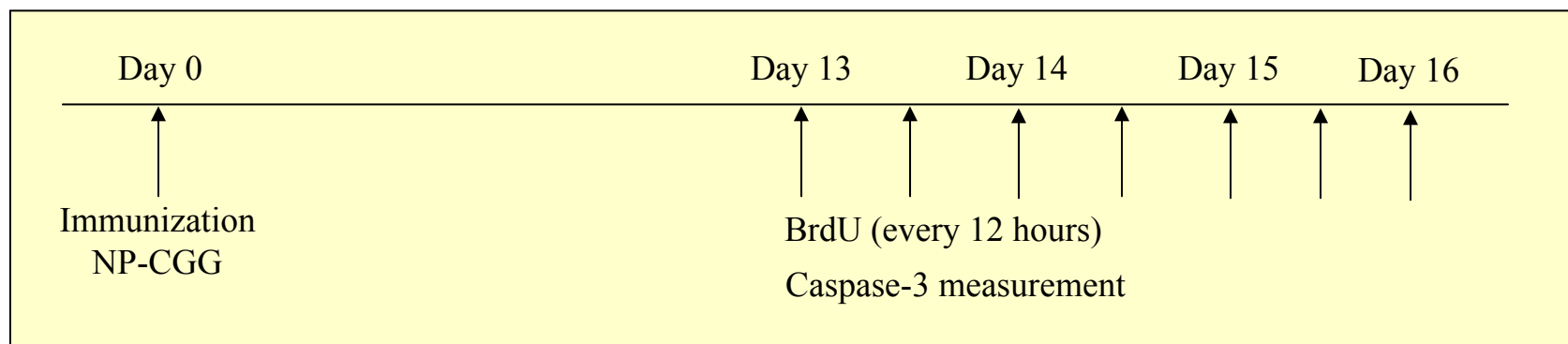
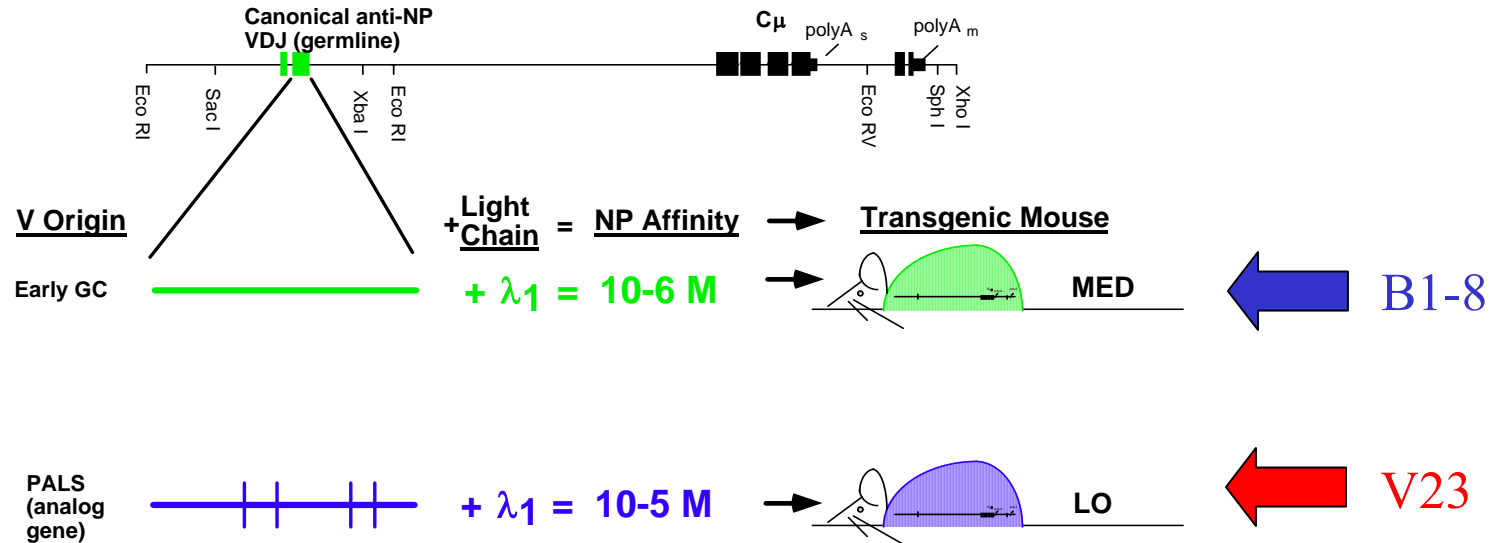
Germinal Centers are Site of Affinity Maturation



Affinity maturation accomplished through somatic hypermutation of B cell receptor, followed by expansion of rare higher-affinity mutants

How does affinity impact cell-fate decisions?

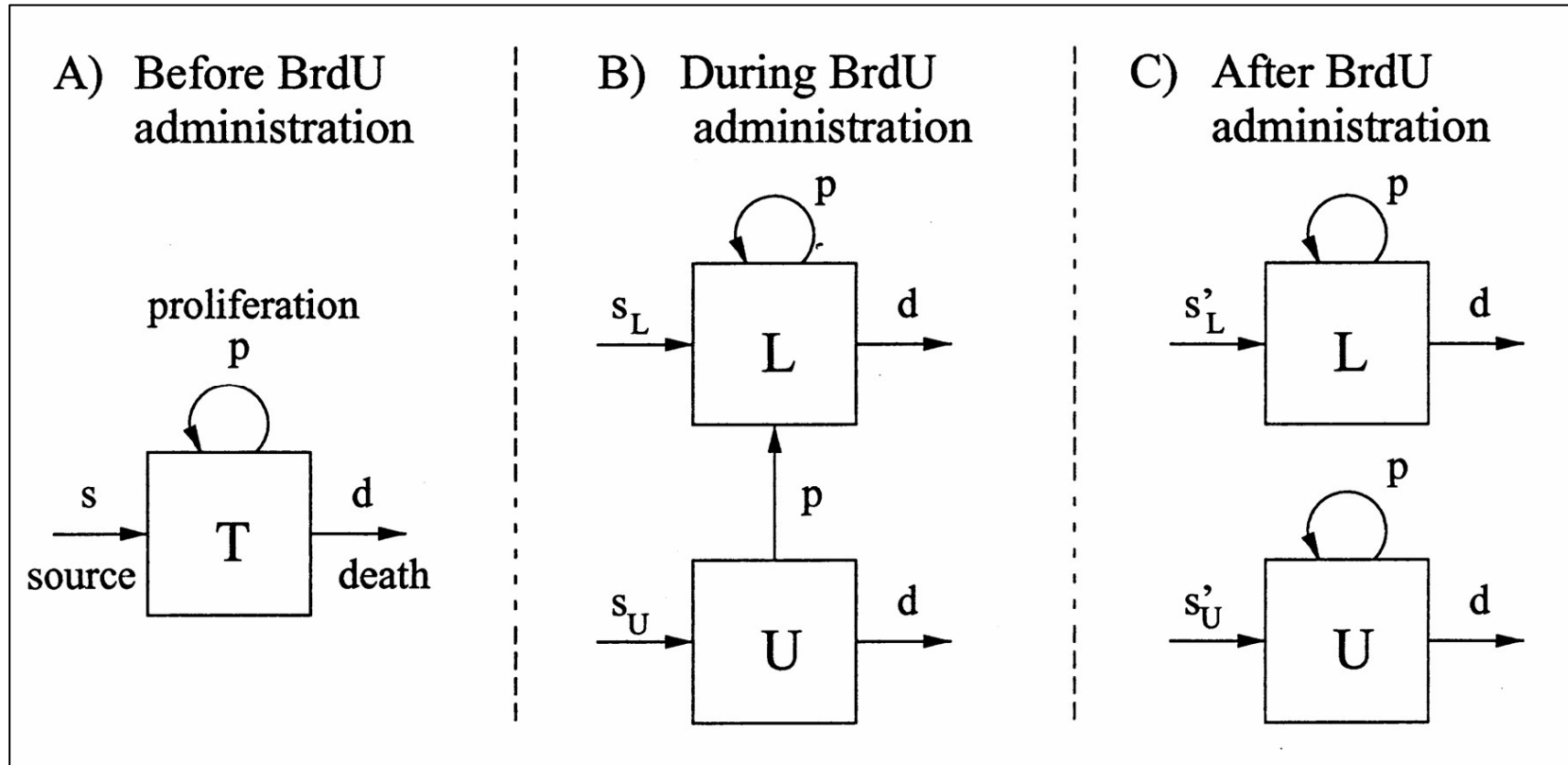
Follow fate of higher and lower affinity B cells using transgenic mice



Is selection driven by a proliferative vs. survival advantage?

Basic Model of BrdU Labeling

Many experiments stop administering label after some time

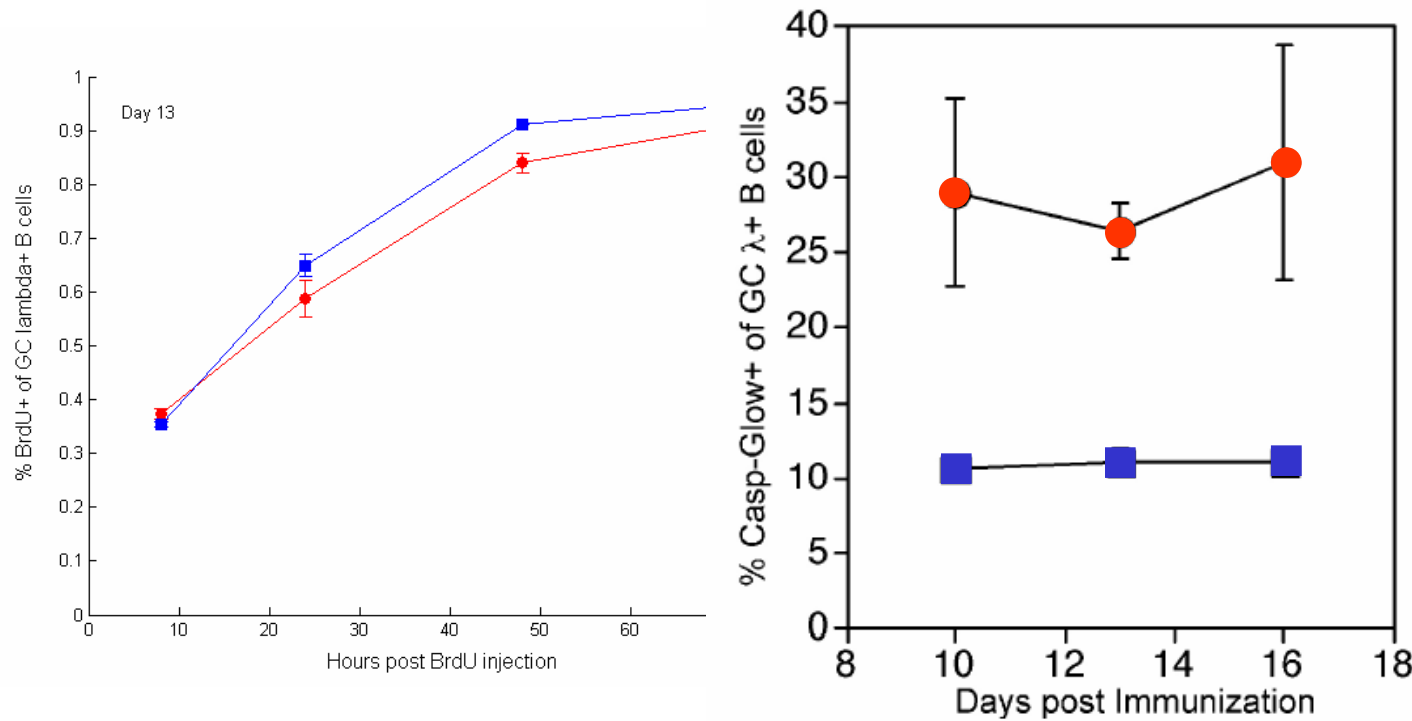


We can express these as sets of ordinary differential equations

How do proliferation and death depend on affinity?

■ Higher Affinity Transgenic (B1-8) ● Lower Affinity Transgenic (V23)

Flow cytometry used to look at antigen-specific germinal center B cells...



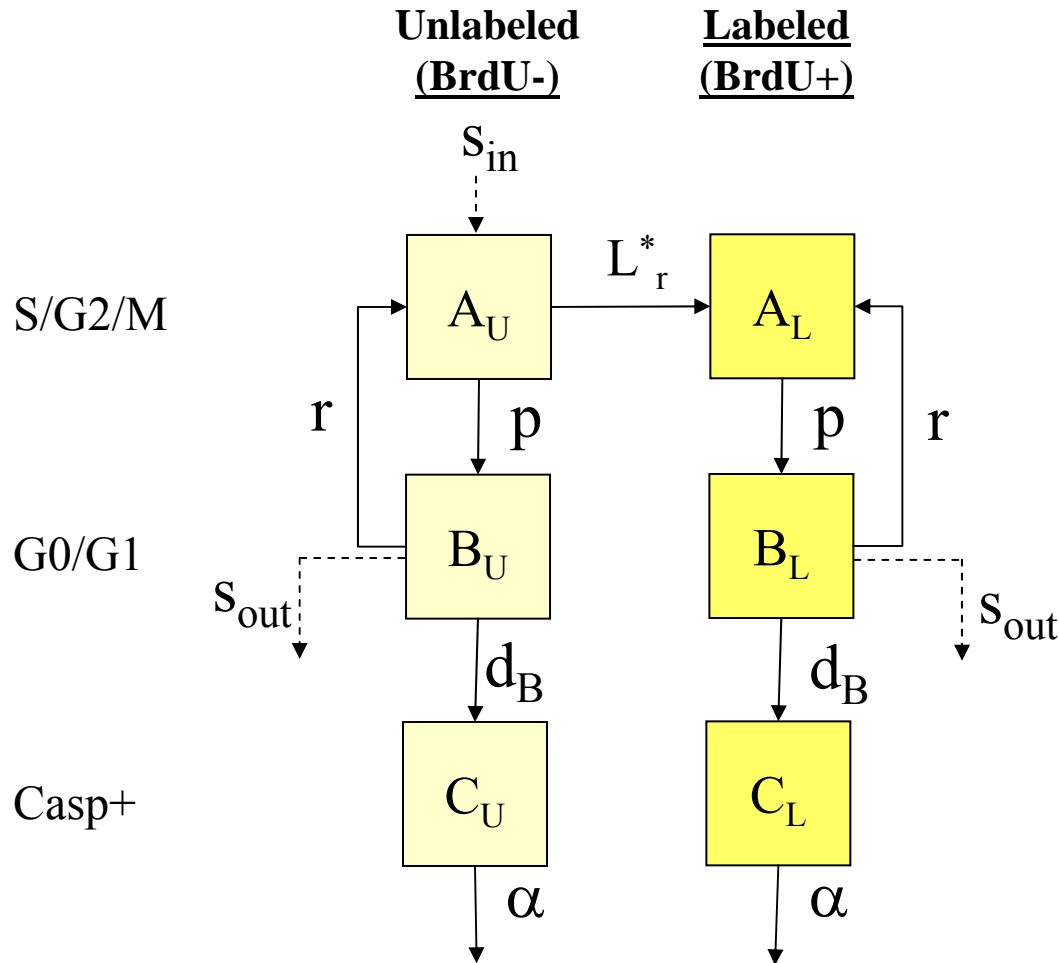
Caspase label tracks dying cells



Updates to basic BrdU model: caspase compartment, BrdU pulse

The ABC Model

A: Dividing (S/G₂/M); **B:** Non-Dividing (G₀/G₁); **C:** CaspGLOW+ cells



$$\frac{dA_U}{dt} = s_{in} + rB_U - (p + d_A + L_r^*)A_U$$

$$\frac{dB_U}{dt} = 2pA_U - (r + d_B + s_{out})B_U$$

$$\frac{dC_U}{dt} = d_A A_U + d_B B_U - \alpha C_U$$

$$\frac{dA_L}{dt} = rB_L + L_r^* A_U - (p + d_A)A_L$$

$$\frac{dB_L}{dt} = 2pA_L - (r + d_B + s_{out})B_L$$

$$\frac{dC_L}{dt} = d_A A_L + d_B B_L - \alpha C_L$$

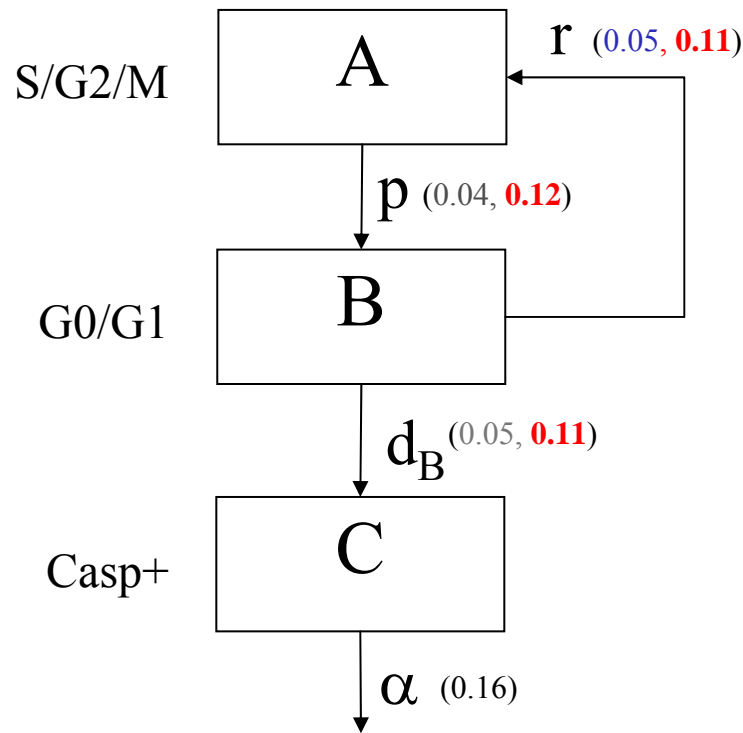
$$L_r^* = \begin{cases} L_r & \text{if } T_i \leq t \leq (T_i + L_t) \\ \text{for some BrdU injection } T_i \\ 0 & \text{otherwise} \end{cases}$$

$$\%BrdU+ = \frac{A_L + B_L + C_L}{A_U + B_U + C_U + A_L + B_L + C_L} \times 100\%$$

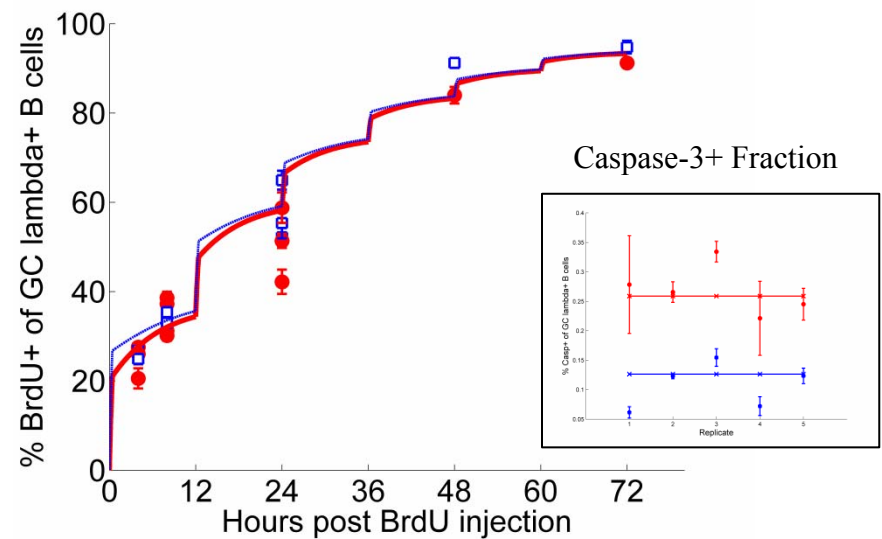
$$\%CaspGLOW+ = \frac{C_U + C_L}{A_U + B_U + C_U + A_L + B_L + C_L} \times 100\%$$

Model estimates proliferation and death rates

■ Higher Affinity Transgenic (B1-8) ● Lower Affinity Transgenic (V23)



The Journal of Immunology
Taking Advantage: High-Affinity B Cells in the Germinal Center Have Lower Death Rates, but Similar Rates of Division, Compared to Low-Affinity Cells¹
 Shannon M. Anderson,^{*} Ashraf Khalil,[†] Mohamed Uduman,^{§§} Uri Hershberg,^{*†}
 Yoram Louzoun,[¶] Ann M. Haberman,[‡] Steven H. Kleinstein,^{§§} and Mark J. Shlomchik^{*†}

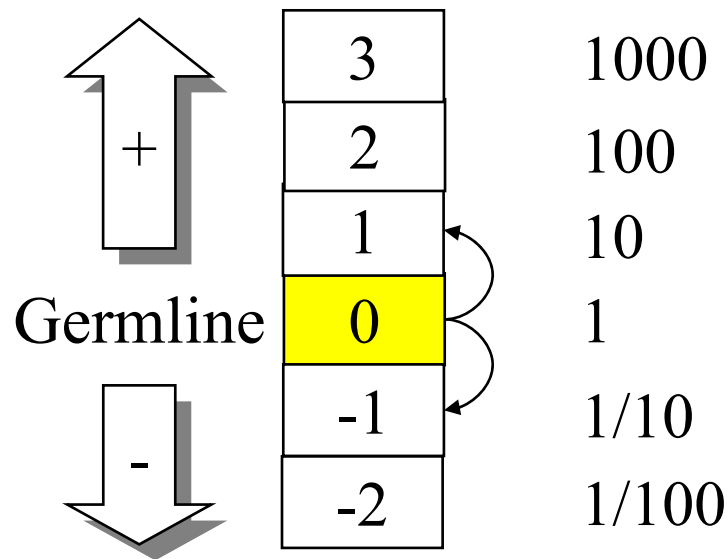


Lower affinity cells have intrinsically **higher death rate**, AND **increased proliferation**

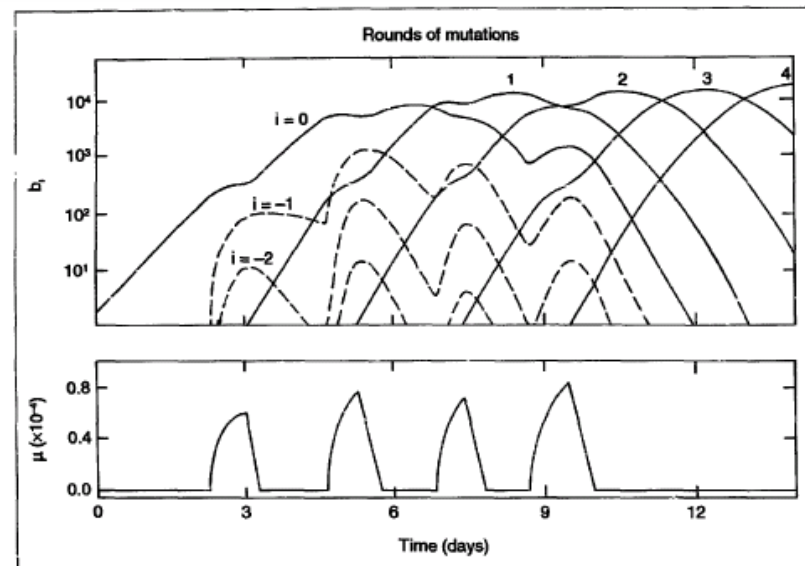
Immune response as optimization problem

Affinity class framework groups B cells with similar on/off-rates

Cyclic re-entry of germinal center B cells and the efficiency of affinity maturation
 Thomas B. Kepler and Alan S. Perelson



$$\frac{db_i}{dt} = b_i \theta_i \left\{ -k_d (1 - h_i) + k_p h_i (2m_u - 1) \right\} + 2k_p \sum_{j \neq i} m_u b_j h_j \theta_j$$



Optimal mutation schedule is phasic (on-off cycles)

But, what should we optimize?

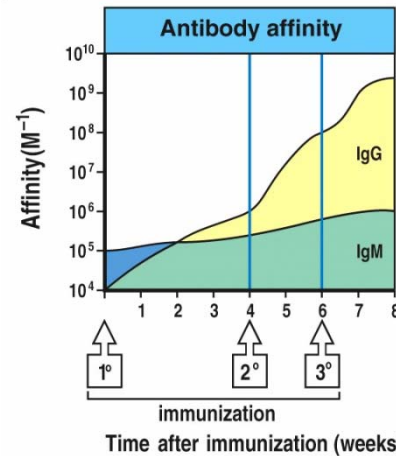


Figure 10-31 Immunobiology, 6/e. (© Garland Science 2005)

Immunology and Cell Biology (1998) 76, 373–381

Theoretical Article

Predicted and inferred waiting times for key mutations in the germinal centre reaction: Evidence for stochasticity in selection

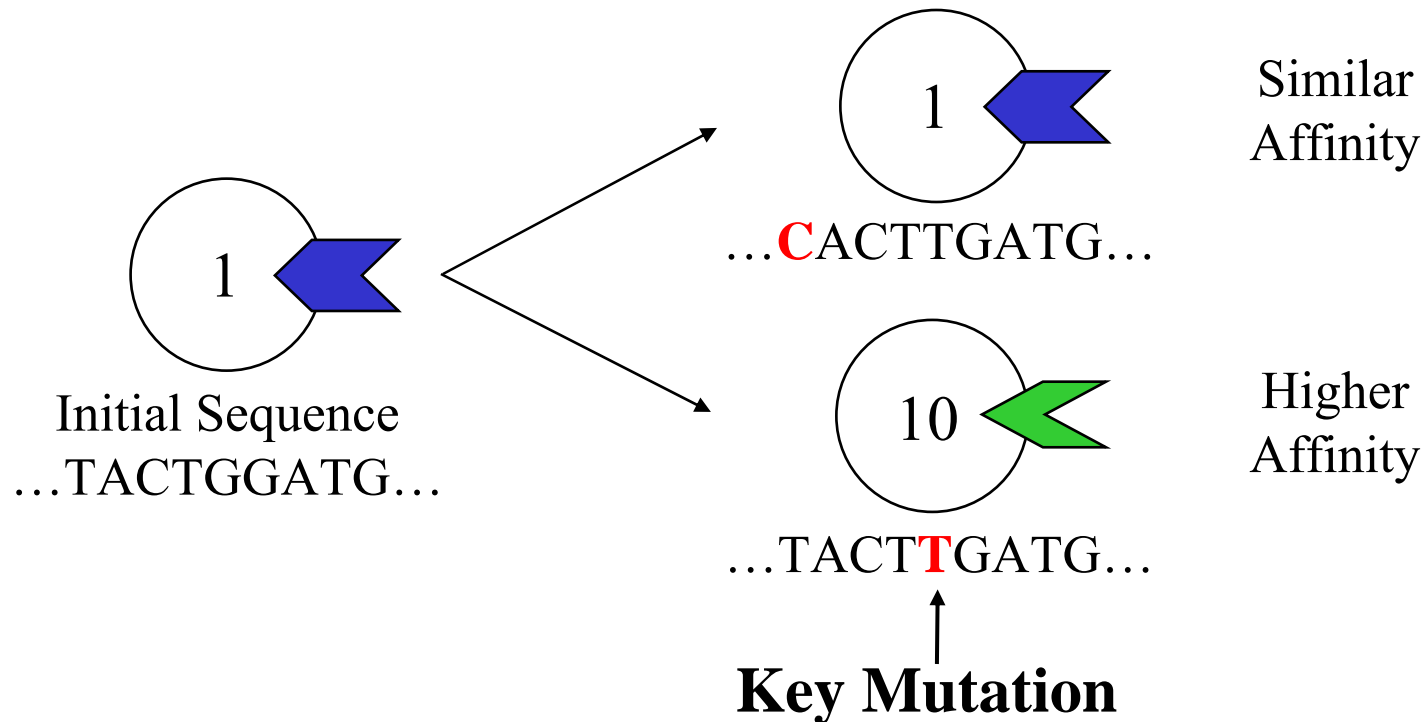
MICHAEL D RADMACHER,¹ GARNETT KELSOE² and THOMAS B KEPLER¹

¹*Biomathematics Graduate Program, Department of Statistics, North Carolina State University, Raleigh, North Carolina,* and ²*Department of Microbiology and Immunology, University of Maryland School of Medicine, Baltimore, Maryland, USA*

How efficient is affinity maturation? Optimal?

Quantitative Affinity Maturation

Consider well-studied antigen NP: (4-hydroxy-3-nitrophenyl)acetyl



Key Mutation increases affinity 10-fold

(majority of high-affinity antibodies observed in the anti-NP response contains this mutation)

Mean waiting time for key mutations

The position 33 mutation, a transversion from G to T in the second nucleotide of the codon, produces a 10-fold increase in binding affinity of the Ig for NP

Number of dividing cells
↓
Division rate
↓
Each division creates two daughter cells
↓

$$\tau = \left(2300 \times 3.43 \times 2 \times 10^{-3} \times 0.19 \times 0.145 \right)^{-1} = 2.3 \text{ days}$$

↑
Average mutation rate
↑
Bias (G → T transversion)
↑
Cold spot (TGT)

Predicted waiting time for key mutations is 2.3 days

Appearance time for key mutations

Experimental sequence data from germinal center microdissections

GC	Strain	Day	Ig sequences*	Position 33 mutations
61AM40	BL/6	8	8	0
61AM41	BL/6	8	10	0
61AM14	BL/6	8	12	0
61AM16	BL/6	8	12	0
L1AB01	BL/6.lpr	10	9	0
L1AB02	BL/6.lpr	10	10	10
L1AB03	BL/6.lpr	10	3	0
L1AB04	BL/6.lpr	10	7	0
61AB08	BL/6	10	4	0
L1AD01	BL/6.lpr	14	12	0
L1AD02	BL/6.lpr	14	11	11
L1AD03	BL/6.lpr	14	10	8
L1AD05	BL/6.lpr	14	11	0
61AD01	BL/6	14	8	3
61AD02	BL/6	14	10	0
61AA02	BL/6	16	8	8

*Sequences are available from EMBL/Gen Bank/DDBJ under accession numbers DS13953 and X67341-X67391.

How does this compare with predicted waiting of 2.3 days

Arrival time of founder key mutant

Two-stage model of B cell mutation and clonal expansion

Stage 0:

Mutation begins ~ day 6.5

Stage I:

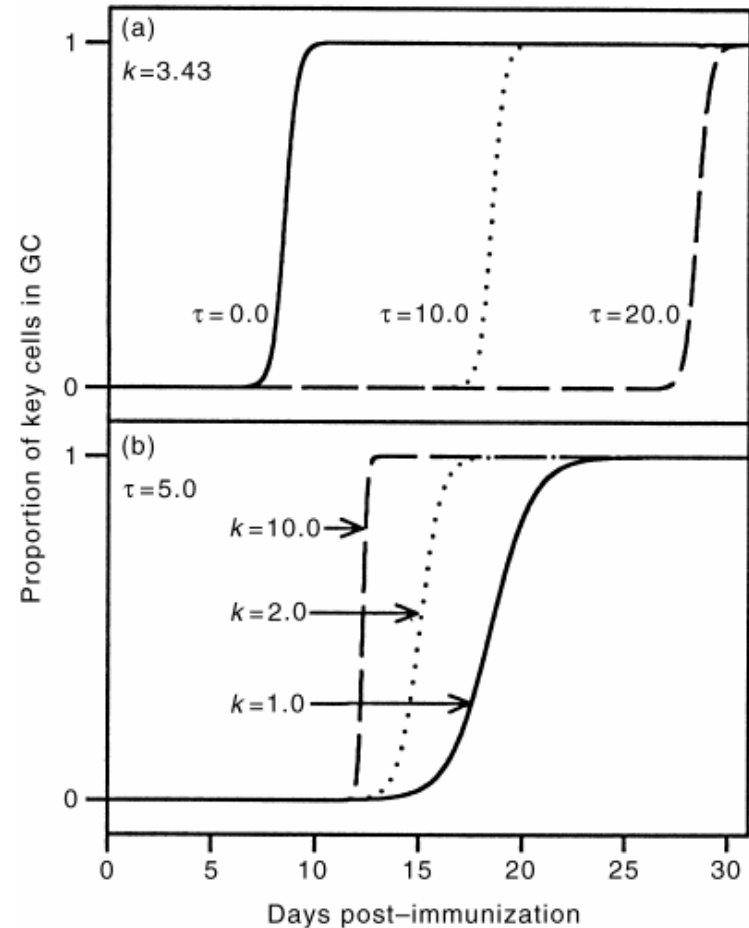
Arrival times are exponentially distributed (τ)

Stage II:

Growth of the key mutant clone is logistic

$$\frac{d\rho}{dt} = k\rho(1 - \rho), \quad \text{for } t > \tau^*$$

↑
Arrival time
of key mutant



Estimate τ and k by fitting to experimental data

Maximum likelihood parameter estimates

Give average appearance time (τ) and proliferation rate (k)...

Observation time

Fraction of key mutants at t_i if first appear at time τ_i^*

Probability to NOT appear by time t_i

$$L = \prod_i \left\{ \int_0^{t_i} [b(x_i | n_i, \rho_i(k, \tau_i^*, t_i)) \cdot f(\tau_i^* | \tau)] d\tau_i^* + I(x_i = 0) \exp\left(-\frac{t_i}{\tau}\right) \right\},$$

Binomial probability (finding x_i key mutants when sampling n_i sequences)

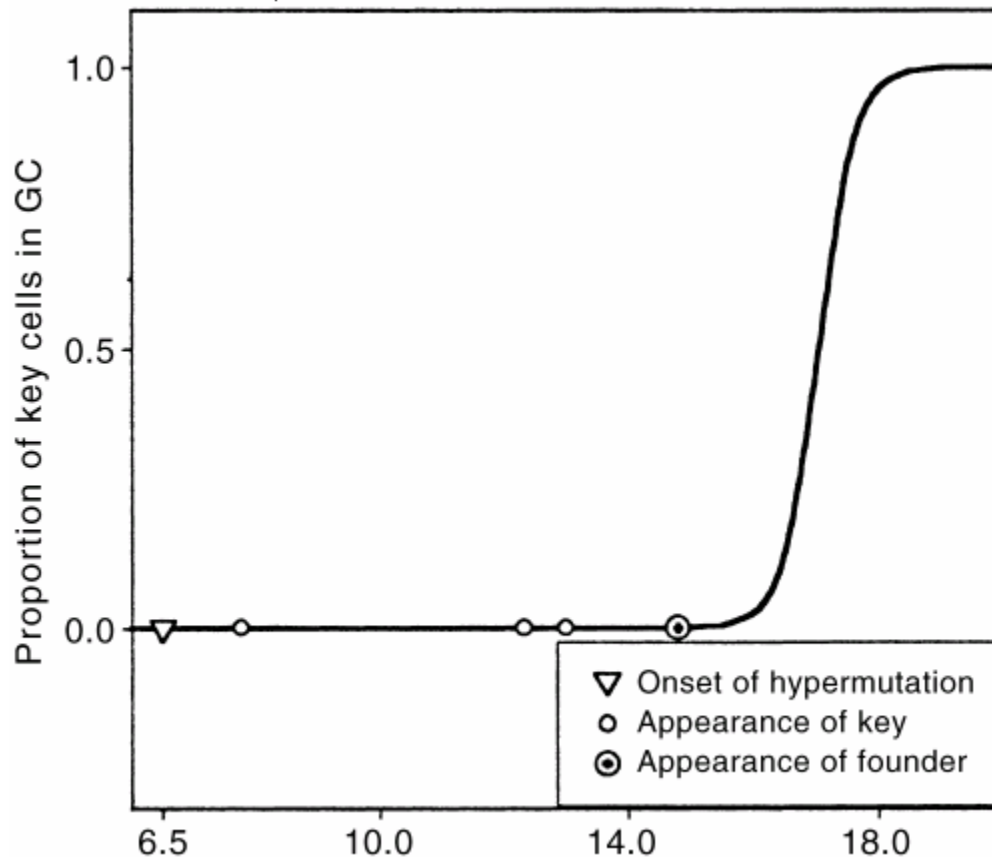
Probability to first appear at time τ_i^*

Appearance time of founder key mutant

Maximize likelihood (L) over τ and k

First key mutant produced earlier than founder

Appearance of founder key mutant is 8.3 days vs. 2.3 days for first key mutant

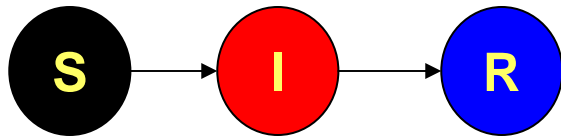


Why?
Stochastic selection
Emigration
Lethal/Blocking mutations

On average, 2.6 key mutants arise that are not perpetuated within the GC before one appears that leads to domination of the GC

The SIR Model of Epidemics

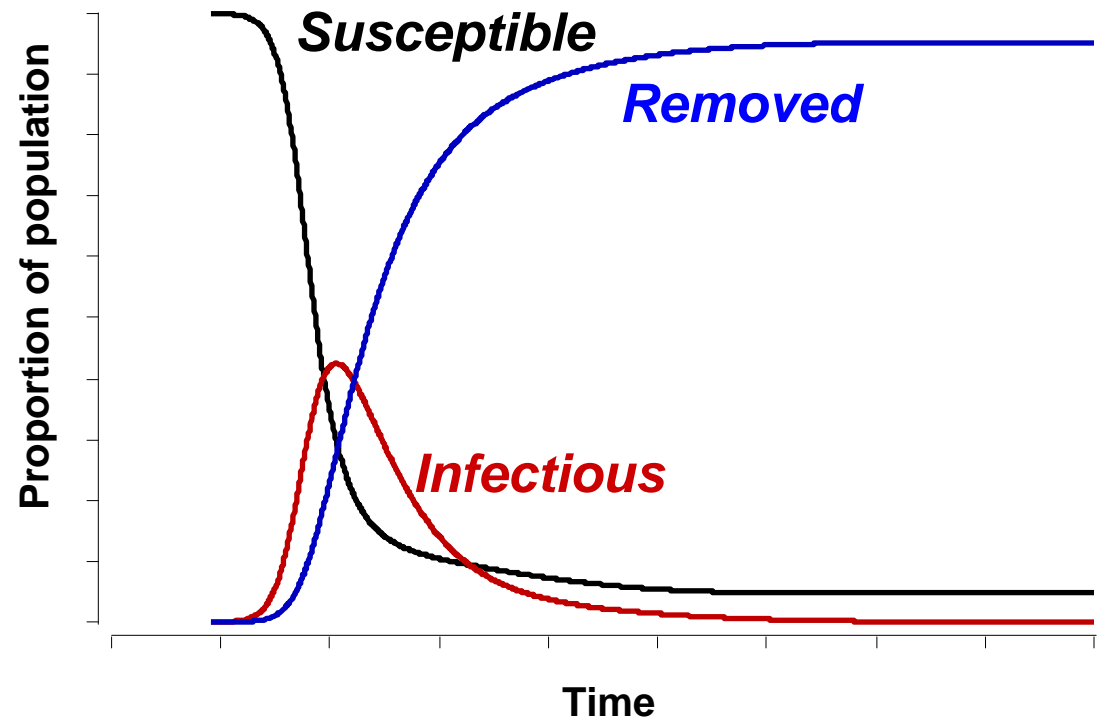
Model for many infectious diseases including measles



$$\frac{dS}{dt} = -\beta SI$$

$$\frac{dI}{dt} = \beta SI - \mu I$$

$$\frac{dR}{dt} = \mu I$$



Other versions allow recovered individual to be re-infected

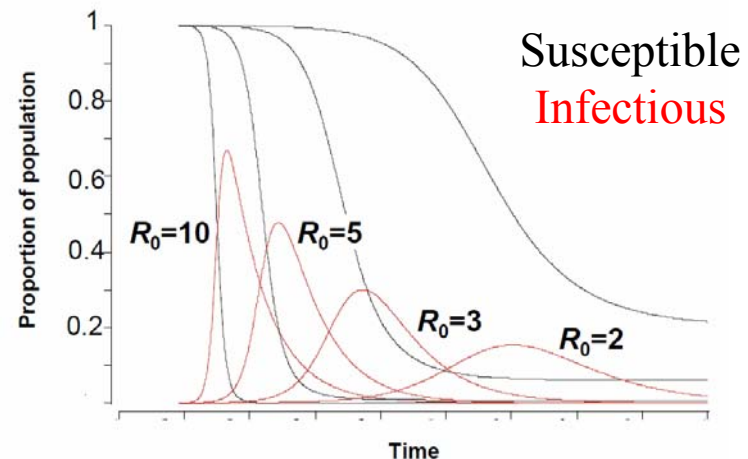
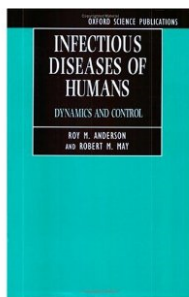
The basic reproductive ratio: R_0

average number of secondary cases caused by an infectious individual in a totally susceptible population

$$R_0 = \frac{\beta}{\mu} \times S(0)$$

$R_0 < 1$: disease dies out

$R_0 > 1$: disease can invade



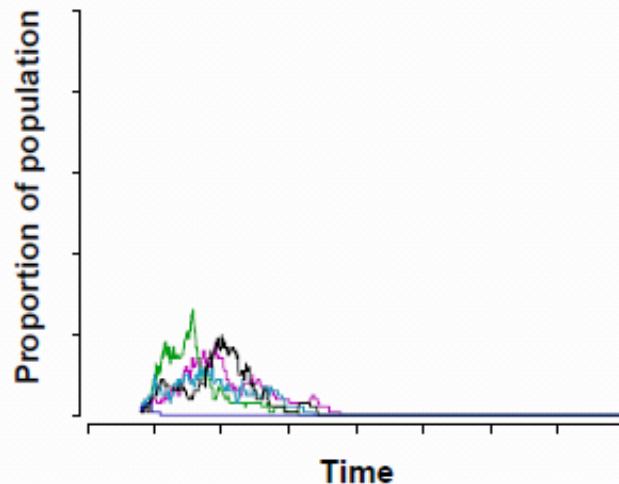
Disease	R_0
AIDS	2 to 5
Smallpox	3 to 5
Measles	16 to 18
Malaria	> 100

R_0 indicates whether population at risk from disease

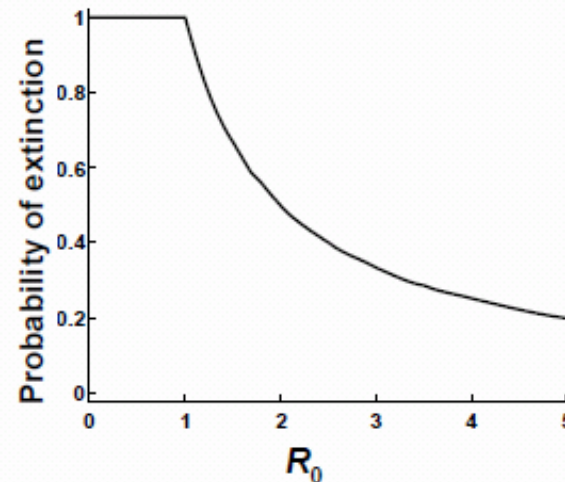
ODEs are deterministic

Predicts epidemic even with non-zero chance that disease dies out

6 stochastic epidemics
with $R_0=3$.



Probability of disease
extinction following
introduction of 1 case.



Stochasticity \rightarrow risk of disease extinction when number of cases is small, even if $R_0 > 1$.

Simulate using stochastic approach – Gillespie Method

Random Numbers

Starting with the same seed will give you equivalent stream

Uniform deviates: [0,1)

Linear congruential generator

$$I_{j+1} = aI_j + c \pmod{m}$$

I_0 is the seed (common to use system clock)

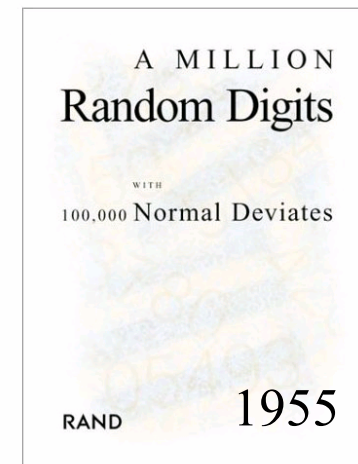
$$I_{j+1} = 3I_j + 7 \pmod{10}$$

Produces: 6,5,2,3

Period: time before stream repeats itself
(maximum m)

Fast, but sequential calls can be correlated, so not used much

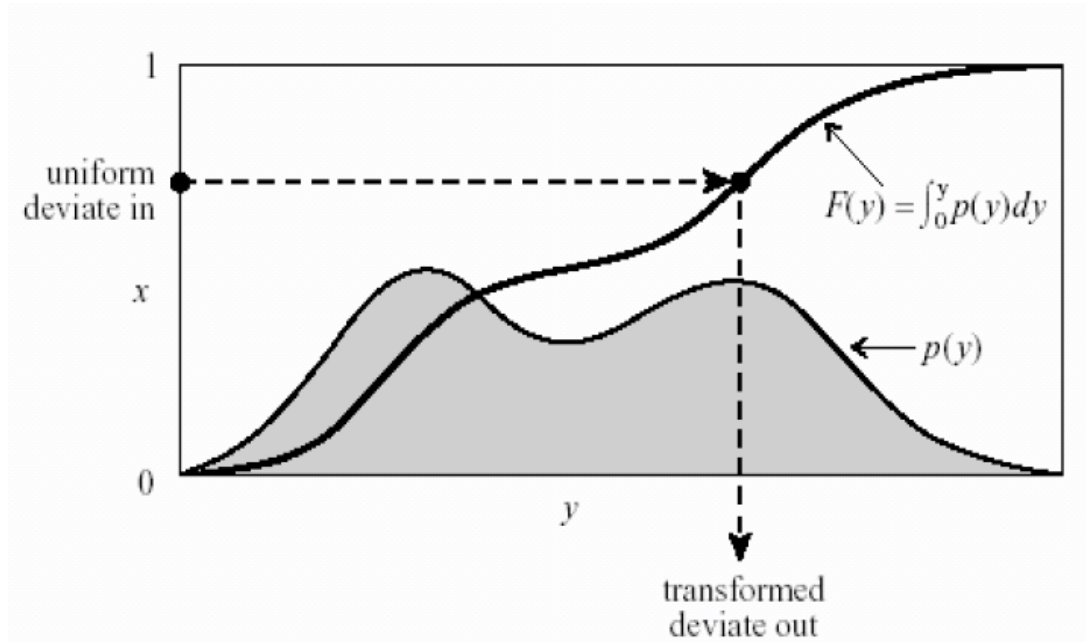
Mersenne Twister
(period $2^{19937}-1$)



Be careful on computer clusters (streams can be correlated)

Simulating from other distributions

Transformation Method: indefinite integral of $p(y)$ must be known and invertible



$$\text{Exponential}(\alpha) = -\frac{1}{\alpha} \ln [\text{Uniform}(0,1)]$$

Transformation to generate exponential distribution (Poisson process)

For more information...

OPEN ACCESS Freely available online

PLoS COMPUTATIONAL BIOLOGY

Message from ISCB

Getting Started in Computational Immunology

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Interdepartmental Program in Computational Biology and Bioinformatics, and Department of Pathology, Yale University School of Medicine, New Haven, Connecticut, United States of America

Feel free to email me with any questions!

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