Protein-Protein Interactions (cont.) & Phosphophorylation

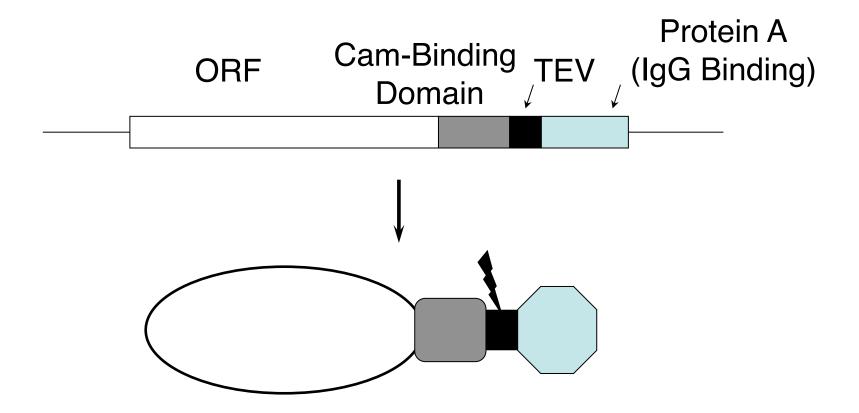
Michael Snyder

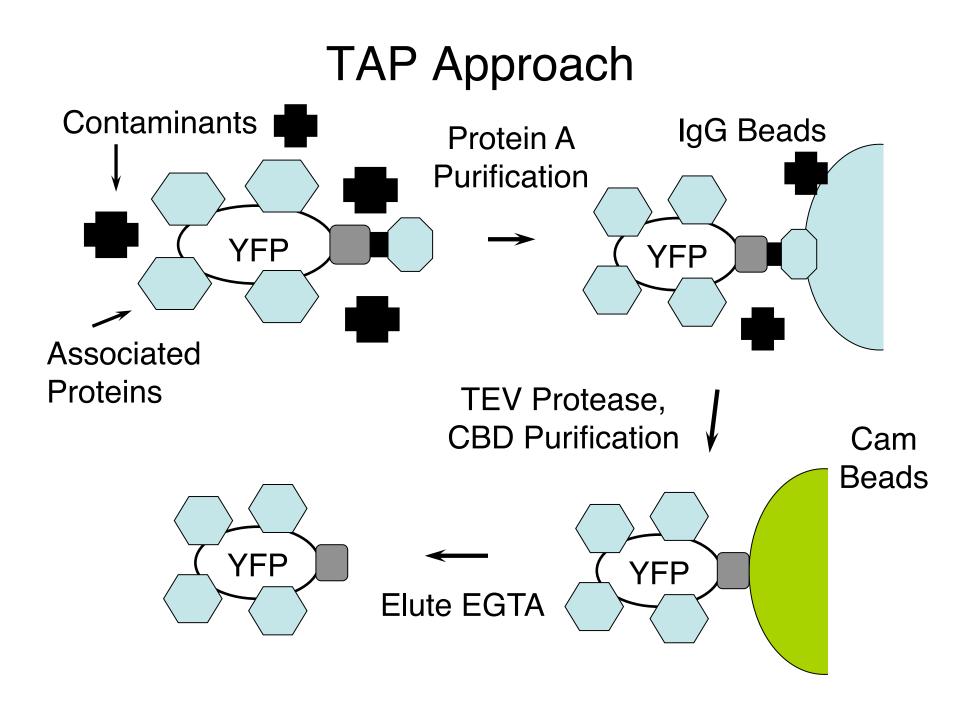
April 8, 2009

Three Methods:

- 1) Two Hybrid
- 2) Complex Analysis: Affinity
 - tagging/Mass Spectrometry
- 3) Protein Chip

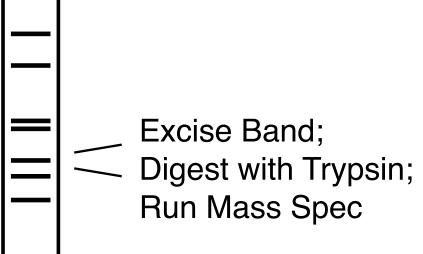
Tandem Affinity Purification (TAP) Tagging

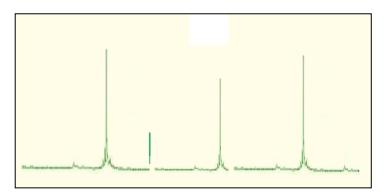




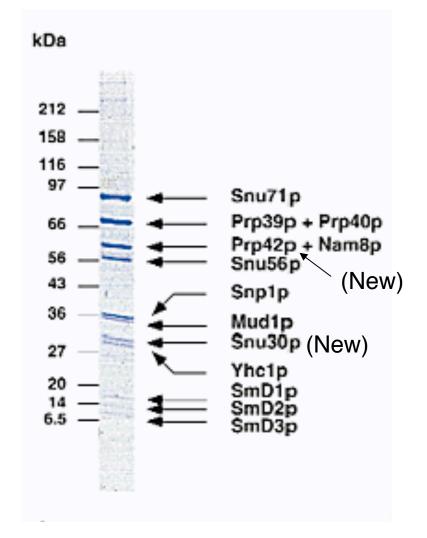
Identify Proteins by Mass Spec

Load on SDS Gel

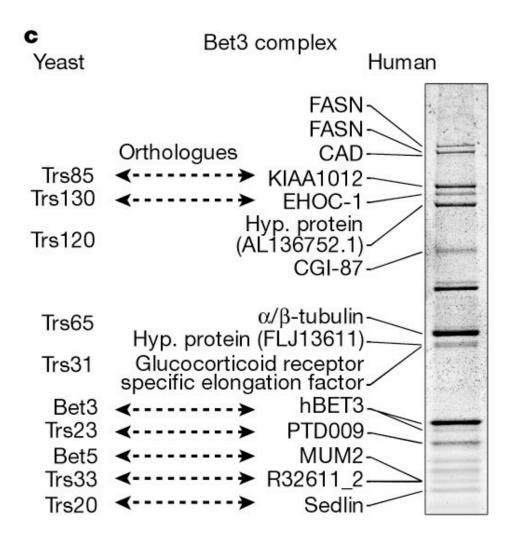




TAP Purification of The U1 Splicing Complex (Snu71p)



Many Complexes Are Conserved



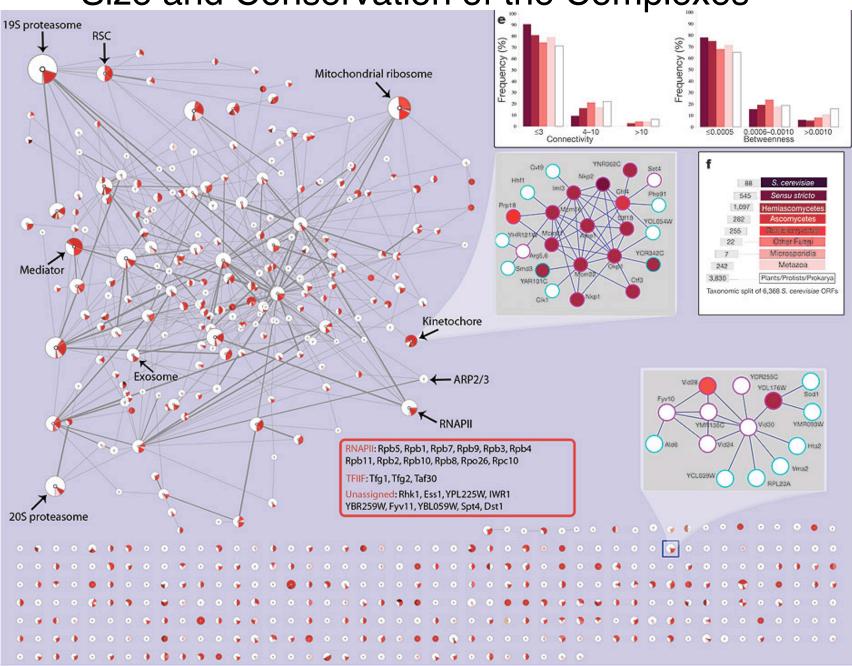
Affinity Purification/Mass Spec Analysis of Complexes - Yeast

4,562 Purifications (Krogan et al. 2002) 2,357 Successful

4,087 Interacting Proteins7,123 Core Interactions (2,708 proteins)14,317 Extended (3,672 proteins)

547 Complexes

Krogan et al. Nature 2006 Vol 440



Size and Conservation of the Complexes

TAP Tag Approach

Advantages

- In Vivo Assay
- Identifies Entire Complex

Disadvantages

- Interactions may be indirect
- Likely to miss some rare components
- Contaminants may copurify

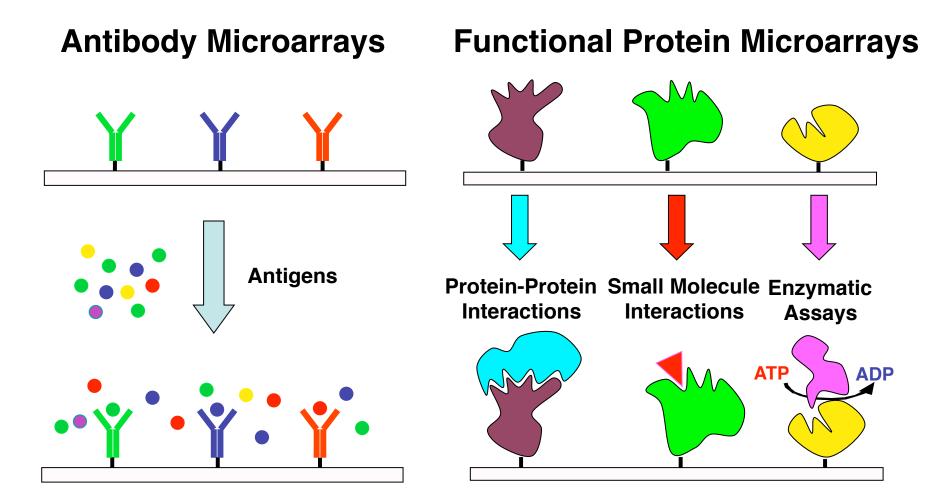
Summary

- Affinity Purification: ~10,000 High Confidence Interactions Among ~3-4000 Proteins
- Two Hybrid: >4,549 Interactions Among 3,278 Proteins
- >20,000 Interactions
- Combining Data = More Accuracy

What is a Protein Microarray?

A high density array containing 100s to many thousands of proteins

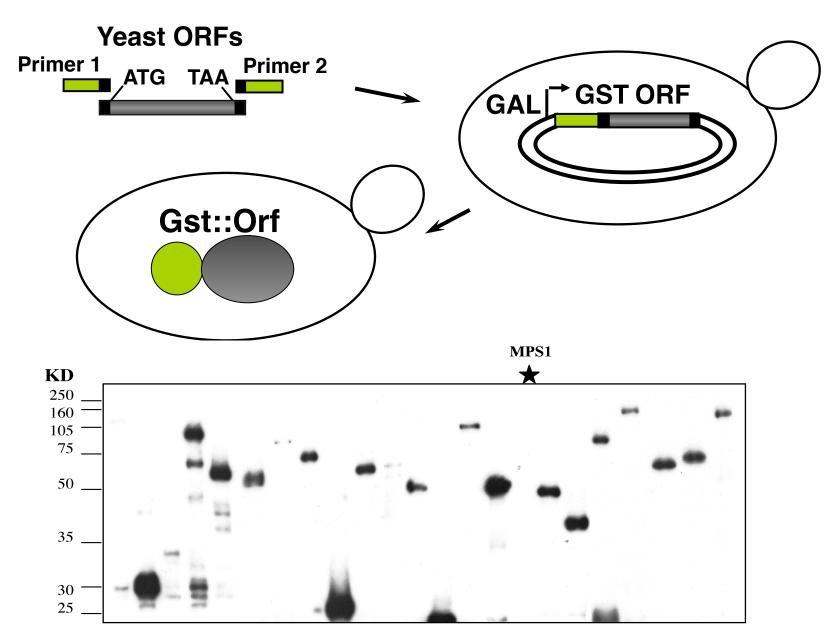
Two Types of Protein Microarrays



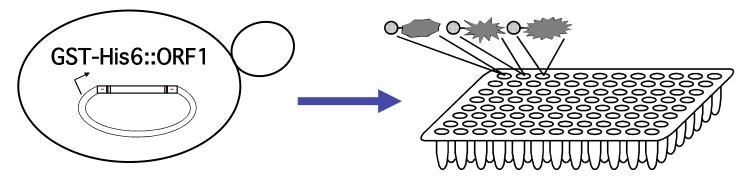
What Is Needed for Preparing and Screening Functional Protein Microarrays?

- A high quality expression library
- Methods for preparing large numbers of proteins
- Methods to array the proteins
- Assays for screening

Cloning & Expression Strategy

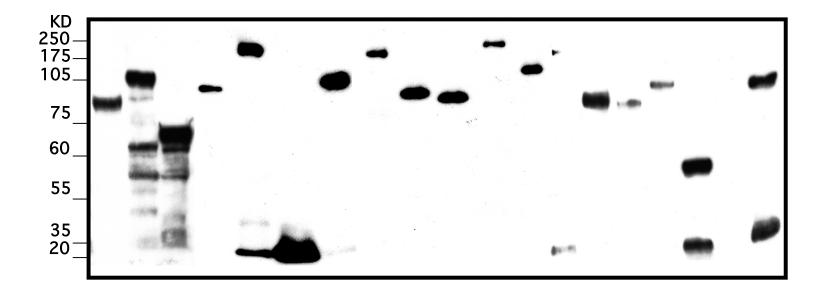


Producing the Yeast Proteome

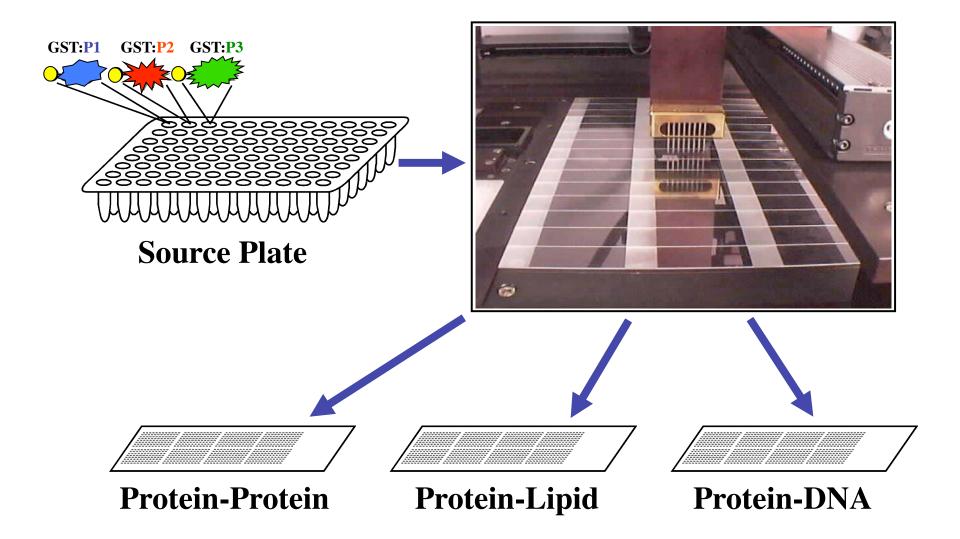


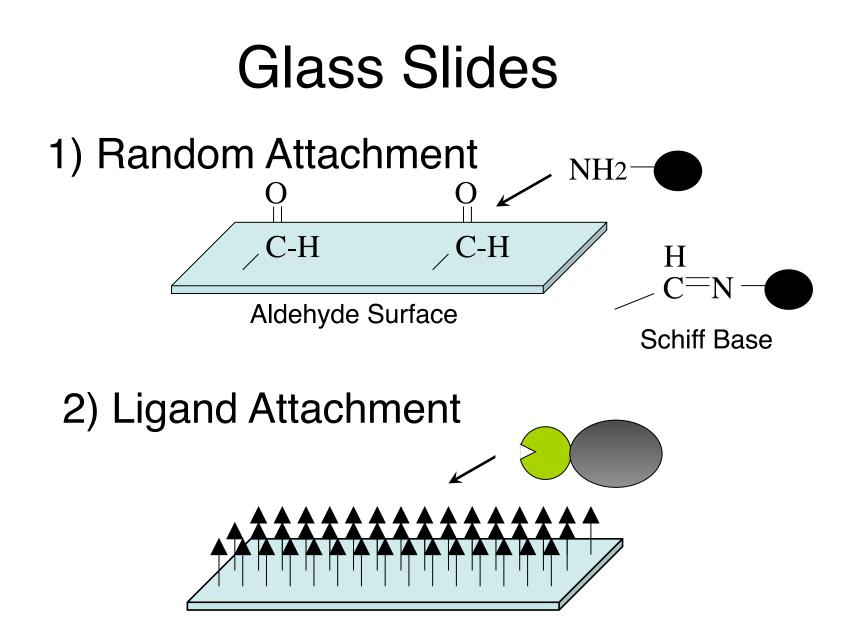
5800 expression clones 93.7%

~80% full-length proteins



Printing the Yeast Proteome





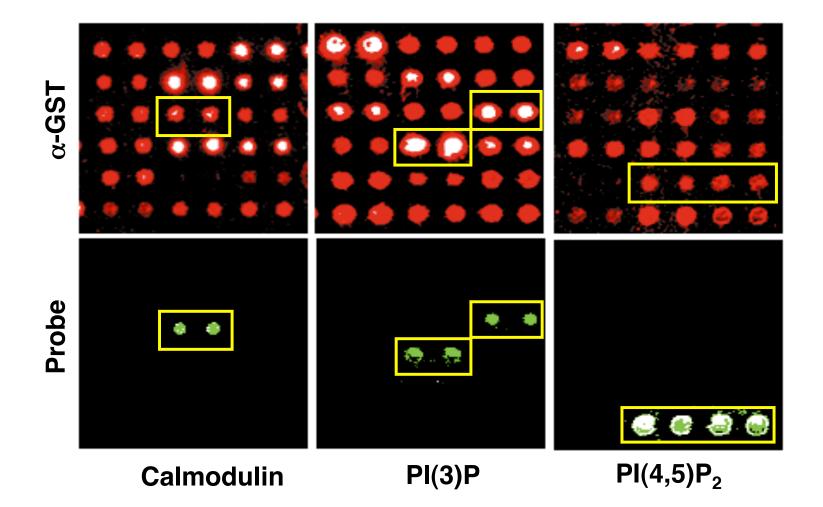
The Yeast Proteome Chip



Many Types of Applications

- Binding Assays
 - Protein-Protein Interactions
 - Protein-Lipid Interactions
 - Nucleic Acids (dsDNA, ssDNA, polyAmRNA)
 - Small Molecule Screens
- Antibody Specificity
- Kinase Substrates and Posttranslational Modifications
- Viral Diagnostics

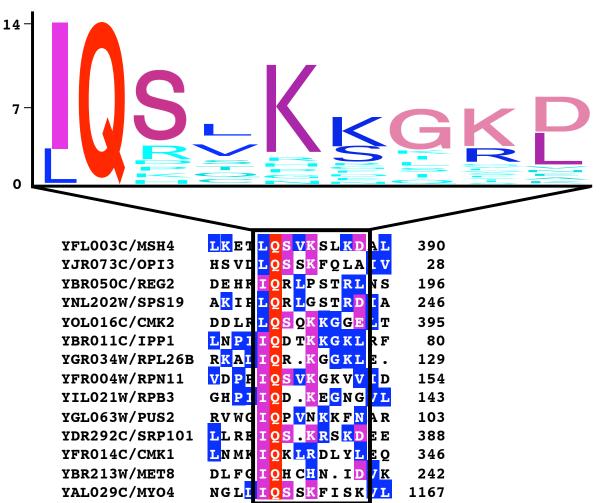
Biochemical Assays on Proteome Chips



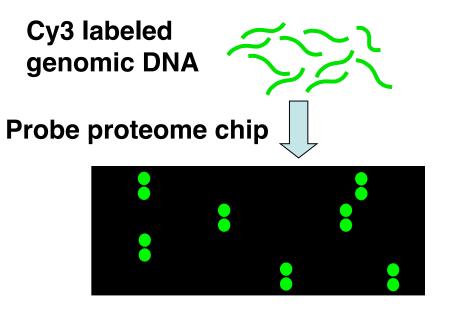
Calmodulin-Binding Proteins

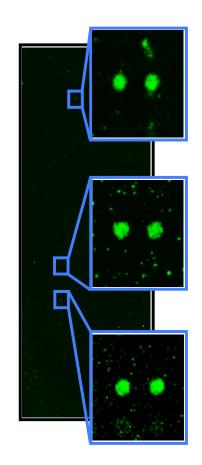
- 12 Known or Suspected Targets
- 33 New Binding Proteins
- Derived New Consensus Binding Site

Potential Calmodulin Binding Motif



Identification of New DNA Binding Activities



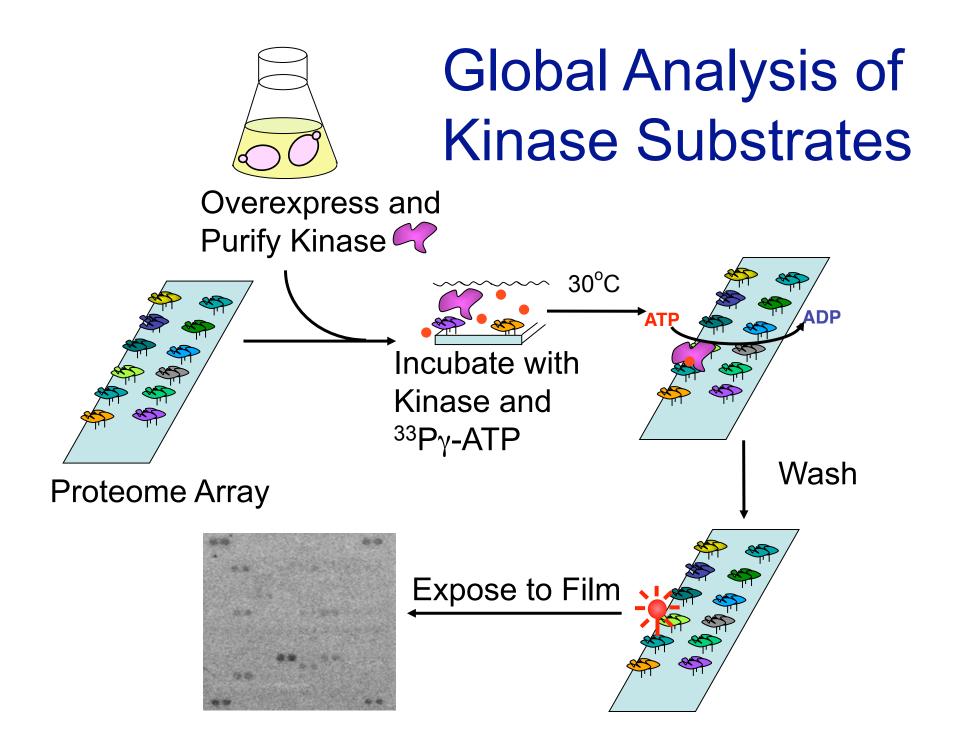


 ~200 bound DNA probe
Found New Activity Arg5,6

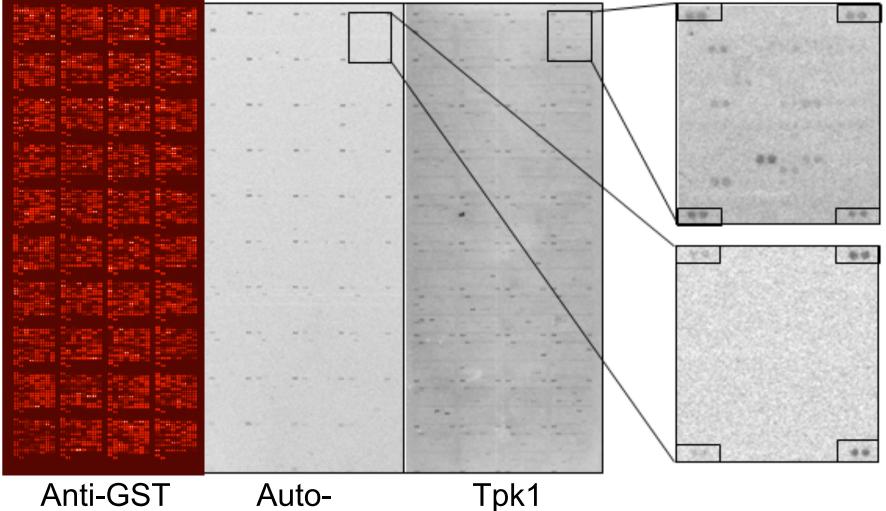
Hall et al. (2004) Science

Yeast Phosphorylation

- 6,000 Proteins 122 Protein Kinase Homologs
 - 14 Uncharacterized
 - 50% Have no known in vivo substrates
 - <160 Known kinase-substrate phosphorylations



Kinase Assays on Protein Chips



ST Auto-Phosphorylation

In Vitro Phosphorylome Summary

AKL1	IKS1	PKH2	STE20
ARK1	IME2	PRK1	SWE1
ATG1	IPL1	PRR1	TOS3
BCK1	IRE1	PRR2	TPK1
CDC15	KCC4	PTK2	TPK2
CDC5	KIN1	RAD53	TPK3
CKA1	KIN2	RCK1	VHS1
CLA4	KIN28	RCK2	YAK1
CMK1	KIN3	RIM11	YCK1
CMK2	KIN4	RIM15	YCK2
CTK1	KIN82	SAT4	YCK3
DBF2	KNS1	SKM1	YDR466W
DUN1	KSP1	SKS1	YGL059W
ELM1	KSS1	SKY1	YGR052W
FUN31	MCK1	SLT2	YKL171W
FUS3	MEK1	SNF1	YMR291W
GIN4	MKK1	SPS1	YOL128C
HAL5	MPS1	SRB10	YOR267C
HRR25	PAK1	SSK22	YPL141C
HSL1	PBS2	STE11	YPL150W
CDC28-CLN2	CDC28-CLB5	PHO85-ALONE	PHO85-PCL1
PHO85-PCL2	PHO85-PCL9	PHO85-PHO80	
RIM15 dead	DBF2 dead	HSL1 dead	RAD53 dead

In Vitro Phosphorylome Summary

- 82 unique kinases and several CDKs with different cyclins (Cdc28; Pho85) for 87 specific hit lists
- 4200 total phosphorylation events on 1325 individual targets
- On average kinase phosphorylated 47 proteins on chip (Range 1- 250)
- 50% of substrates were phosphorylated by only one kinase
- Identified at least 21 known kinase-substrate phosphorylations

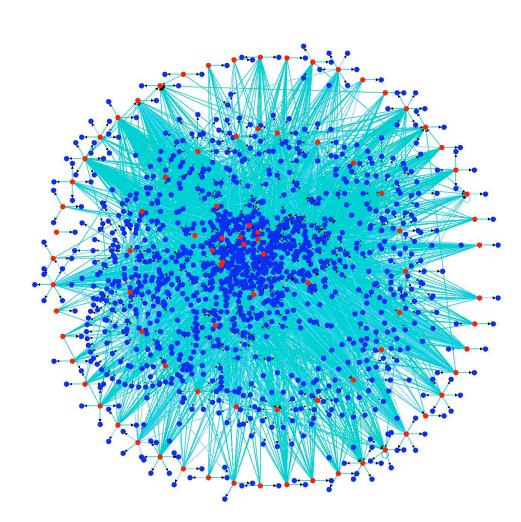
Substrate List Comparison



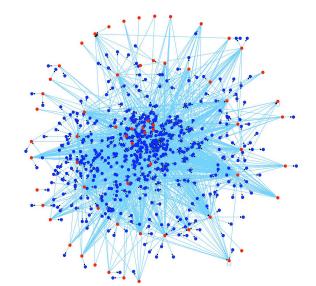
TPK1, PKA overlapped very well

Pcl9, Pho80 overlapped very well

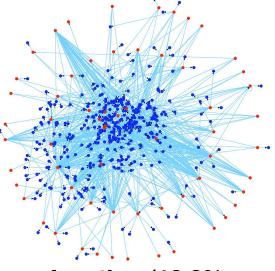
Phosphorylome Network



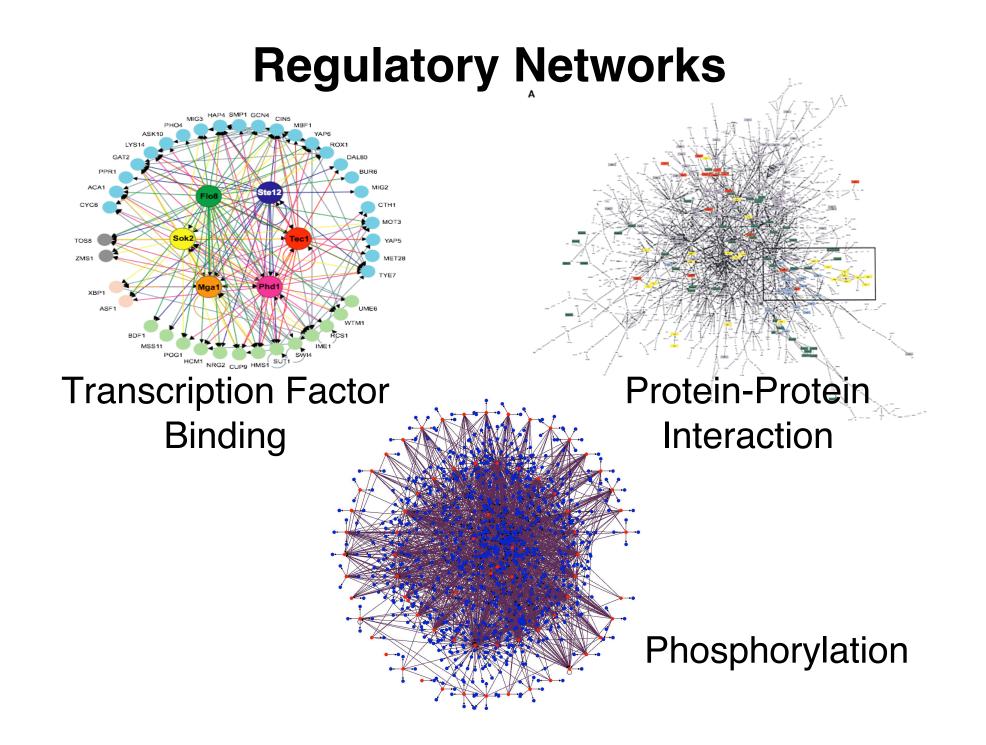
4200 Phosphorylations, 1325 Proteins

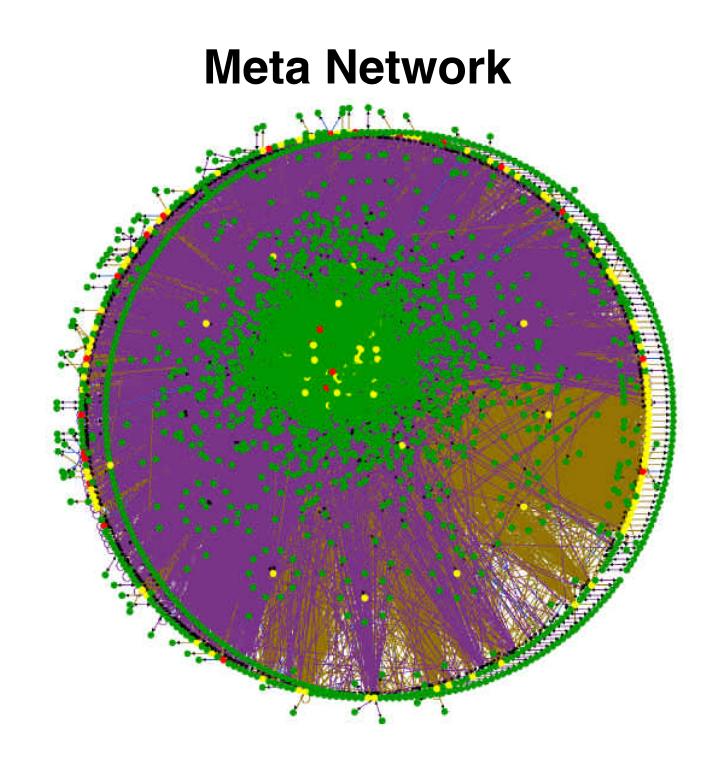


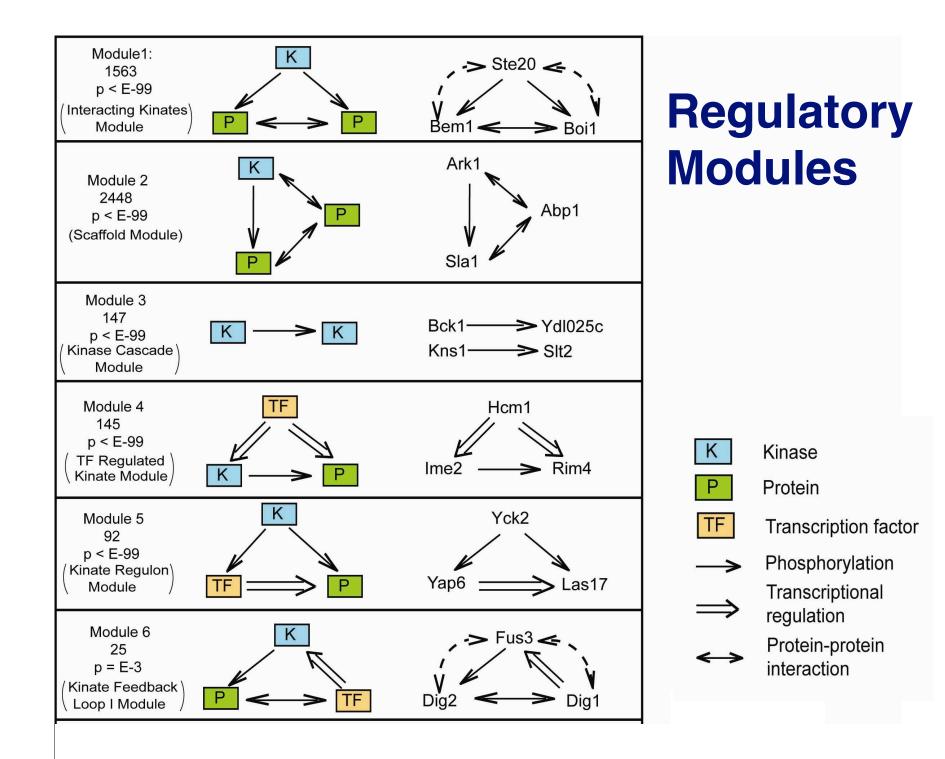
Same localization (33%; p E<-99)



Same function (18.3%; p <E-99)







Advantages of Protein Chips

- Can screen many proteins simultaneously
- Small amounts of proteins and reagents
- High throughput
- Diverse applications-biochemical assays, posttranslational modifications, small molecule screening

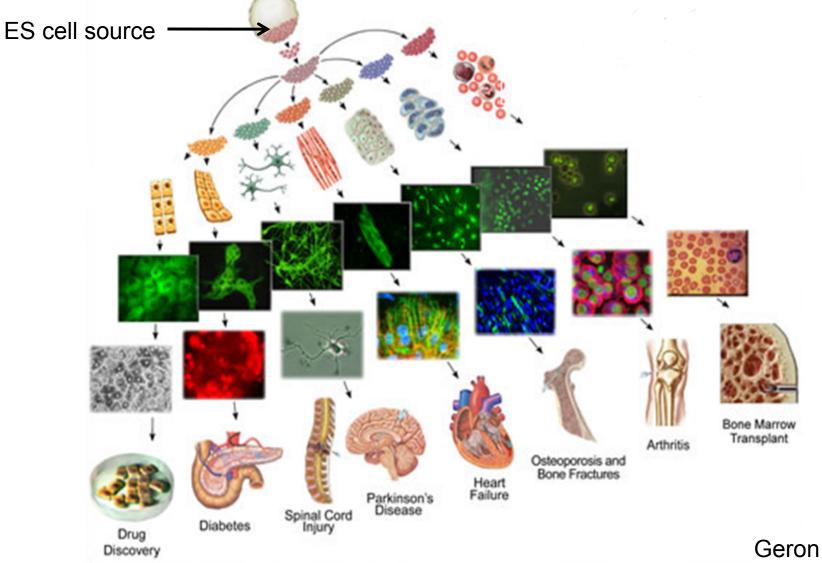
Disadvantage

• In Vitro Assay - Must validate In vivo

Many Transcription Factors Remain Uncharacterized

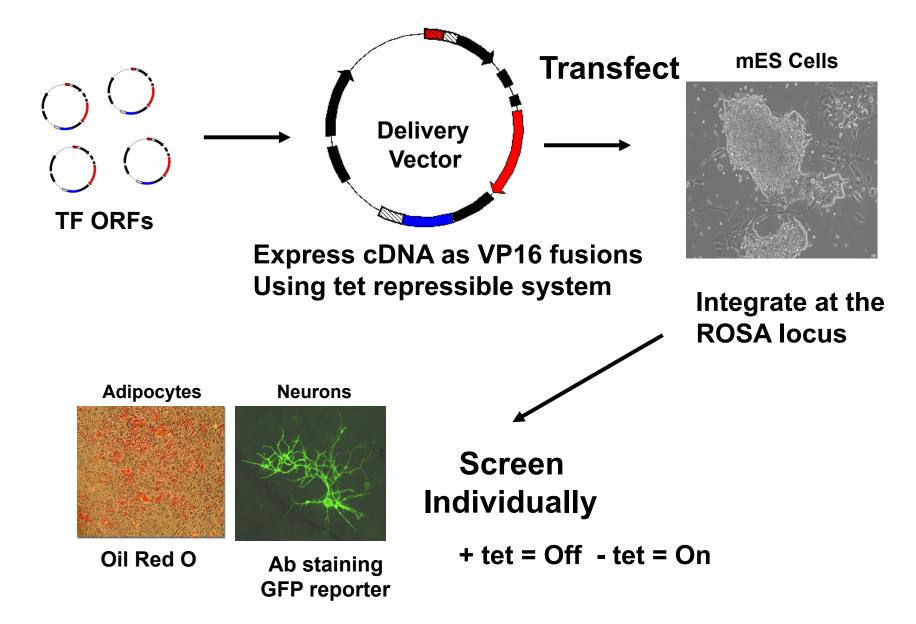
- * 1400-1700 human transcription factors
- Methods of studying TF function in differentiation
 - Map binding sites
 - Tissue/cell localization
 - Knock down
 - Ectopic expression
- Ectopically expressed TFs with the ability to activate differentiation programs
 - MyoD (muscle), PPARgamma (fat), Sox 5, 6, 9 (cartilage)

Embryonic Stem Cells are a 'Blank Slate' for Studying Gene Function



Geron Corp.

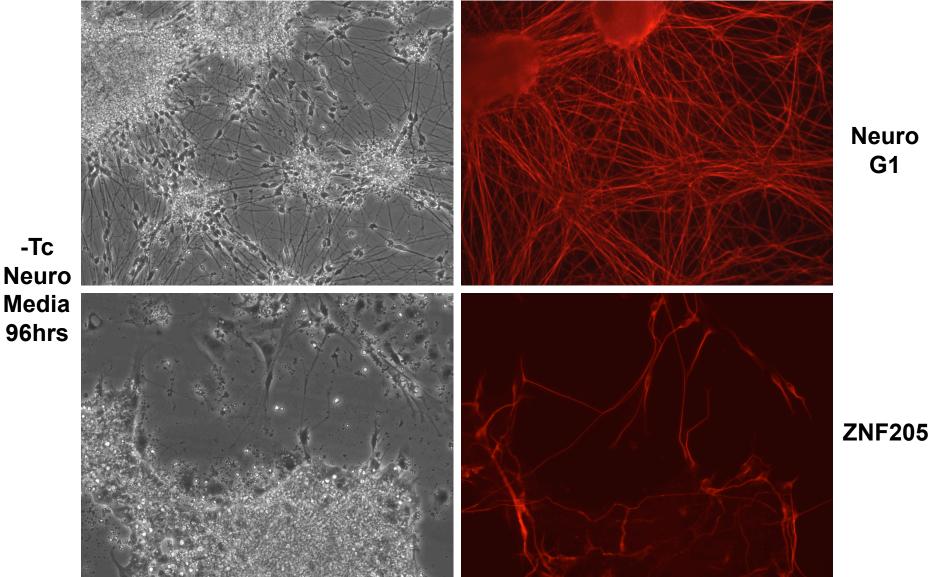
Screen 700 Human TFs in mES Cells



24 TFs Induce Differentiation

Phase

anti-beta III tubilin

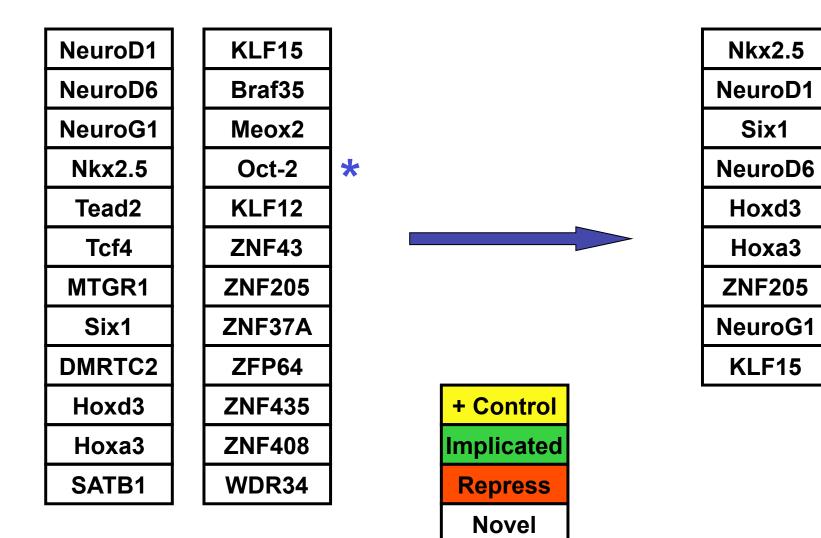


Neuro

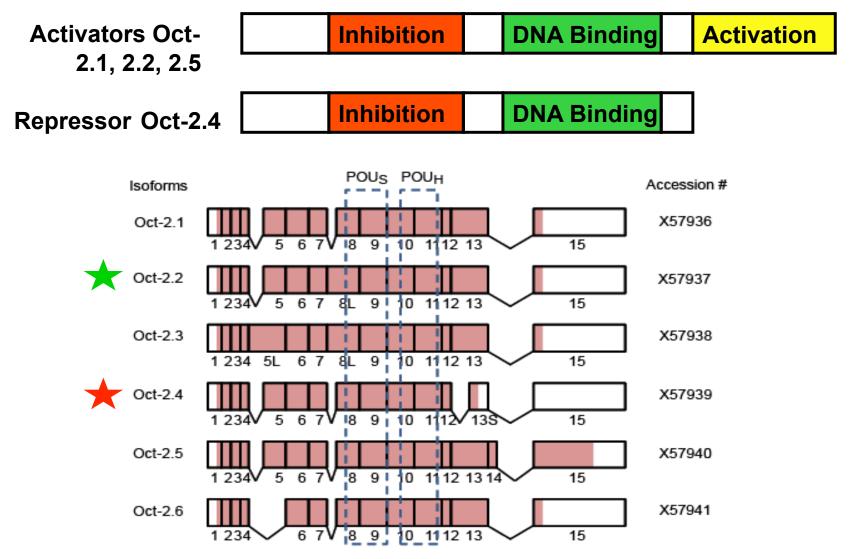
24 TFs Induce Neural Differentiation

Hits with VP16

Hits without VP16



Oct-2 Many Isoforms



Adapted from Dong et al., 2007

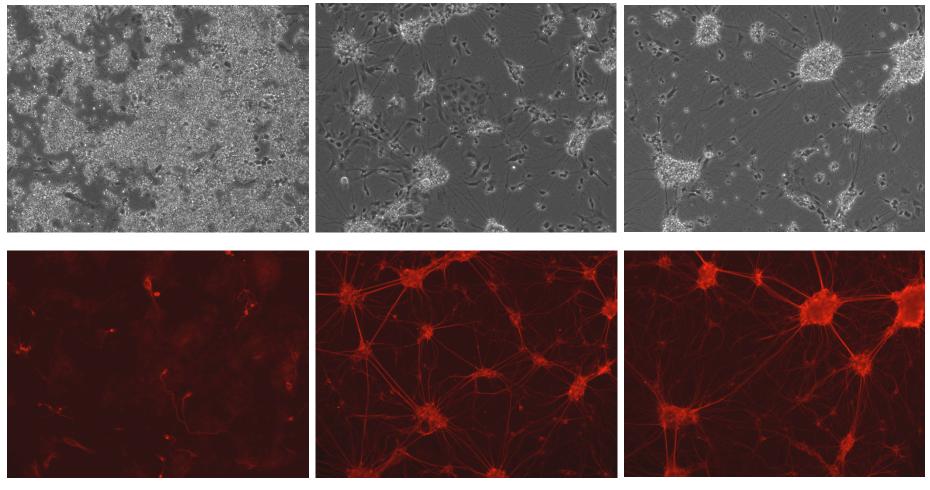
Oct-2.4 Inhibits Neuronal Differentiation

EBs dissociated and plated onto poly-ornithine/laminin coated plates

Oct-2.4

EBRTcH3 (WT)

Oct-2.4::VP16

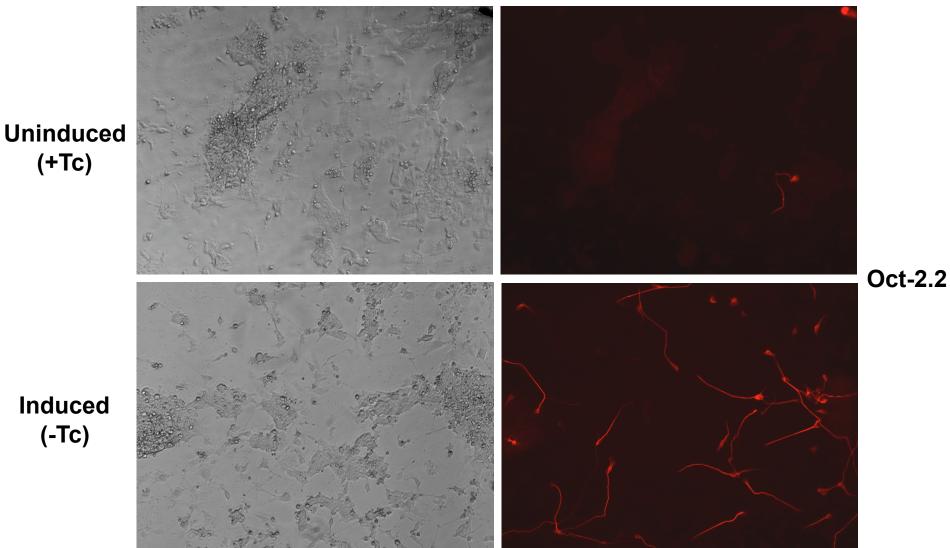


anti-βlll tubulin

Oct-2.2 Isoform Induces Differentiation

Phase

anti-βlll tubulin



(+Tc)

Oct-2 Differentiation Model

