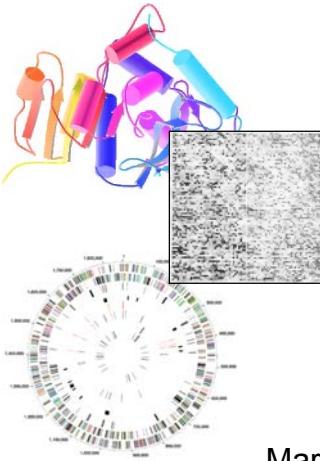


BIOINFORMATICS

Structures



Mark Gerstein, Yale University
bioinfo.mbb.yale.edu/mgb452a

Contents: Structures

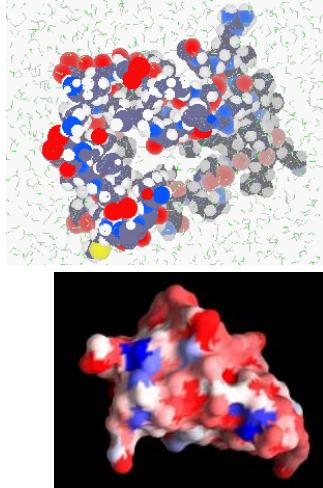
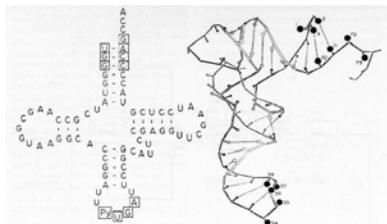
- What Structures Look Like?
- Structural Alignment by Iterated Dynamic Programming
 - ◊ RMS Superposition
 - ◊ Rotating and Translating Structures
- Scoring Structural Similarity
- Other Aspects of Structural Alignment
 - ◊ Distance Matrix based methods
- Elaborating structures
 - ◊ Surfaces and volumes

Molecular Biology Information: Macromolecular Structure

- DNA/RNA/Protein

 - ◊ Almost all protein

(RNA Adapted From D Sol Web Page.
Right hand Top Protein from M Levitt web page)



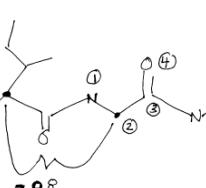
3 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

Molecular Biology Information: Protein Structure Details

- Statistics on Number of XYZ triplets

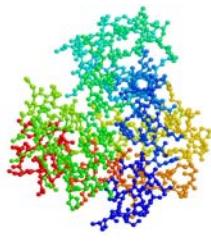
 - ◊ 200 residues/domain → 200 CA atoms, separated by 3.8 Å
 - ◊ Avg. Residue is Leu: 4 backbone atoms + 4 sidechain atoms, 150 cubic Å
 - => ~1500 xyz triplets (=8x200) per protein domain
 - ◊ 10 K known domain, ~300 folds

ATOM	1	C	ACE	0	9.401	30.166	60.595	1.00	49.88	IGKY	67
ATOM	2	O	ACE	0	10.432	30.832	60.722	1.00	50.35	IGKY	68
ATOM	3	CH3	ACE	0	8.876	29.767	59.226	1.00	50.04	IGKY	69
ATOM	4	N	SER	1	8.753	29.755	61.685	1.00	49.13	IGKY	70
ATOM	5	CA	SER	1	9.242	30.205	62.974	1.00	46.62	IGKY	71
ATOM	6	C	SER	1	10.453	29.501	63.579	1.00	41.99	IGKY	72
ATOM	7	O	SER	1	10.593	29.607	64.814	1.00	43.24	IGKY	73
ATOM	8	CB	SER	1	8.052	30.189	63.974	1.00	53.00	IGKY	74
ATOM	9	OB	SER	1	7.294	31.409	63.930	1.00	57.79	IGKY	75
ATOM	10	N	ARG	2	11.360	28.811	62.827	1.00	36.48	IGKY	76
ATOM	11	CA	ARG	2	12.548	28.316	63.532	1.00	30.20	IGKY	77
ATOM	12	C	ARG	2	13.502	29.501	63.500	1.00	25.54	IGKY	78
ATOM	1444	CB	LYS	186	13.836	22.263	57.567	1.00	55.06	IGKY1510	
ATOM	1445	CG	LYS	186	12.422	22.452	58.180	1.00	53.45	IGKY1511	
ATOM	1446	CD	LYS	186	11.531	21.198	58.185	1.00	49.88	IGKY1512	
ATOM	1447	CE	LYS	186	11.452	20.402	56.860	1.00	48.15	IGKY1513	
ATOM	1448	NZ	LYS	186	10.735	21.104	55.811	1.00	48.41	IGKY1514	
ATOM	1449	OXT	LYS	186	16.887	23.841	56.647	1.00	62.94	IGKY1515	
TER	1450		LYS	186							IGKY1516

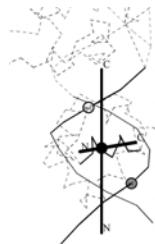


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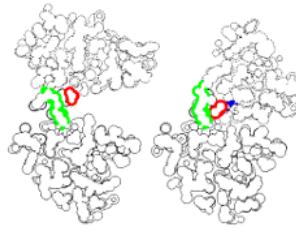
Other Aspects of Structure, Besides just Comparing Atom Positions



Atom
Position,
XYZ triplets



Lines, Axes,
Angles

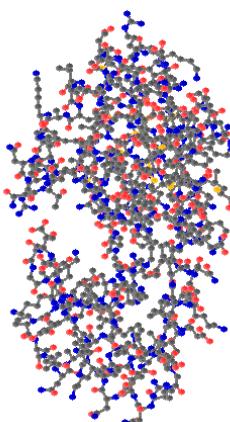


Surfaces, Volumes

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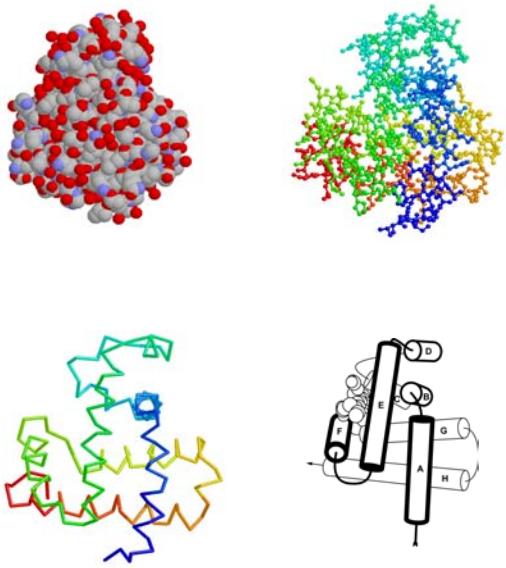
What is Protein Geometry?

- Coordinates (X, Y, Z's)
- Derivative Concepts
 - ◊ Distance, Surface Area, Volume, Cavity, Groove, Axes, Angle, &c
- Relation to
 - ◊ Function, Energies ($E(x)$), Dynamics (dx/dt)

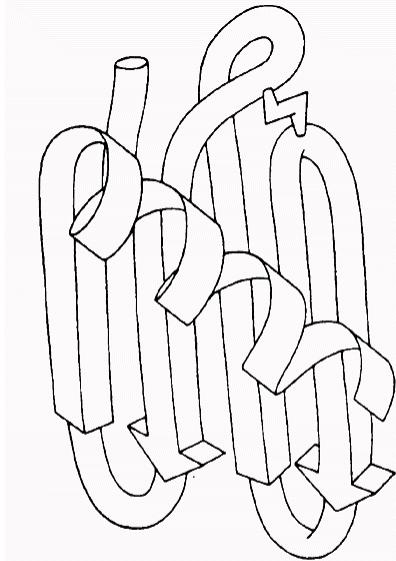


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Depicting
Protein
Structure:
Sperm
Whale
Myoglobin



7 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu



Incredulase

Incredulase

J.S. Richardson and D.C. Richardson, "Some design principles: Betabellin", in
D.L. Oxender and C.F. Fox (Eds.), "Protein Engineering", Alan R. Liss, 1987, p.
149-163

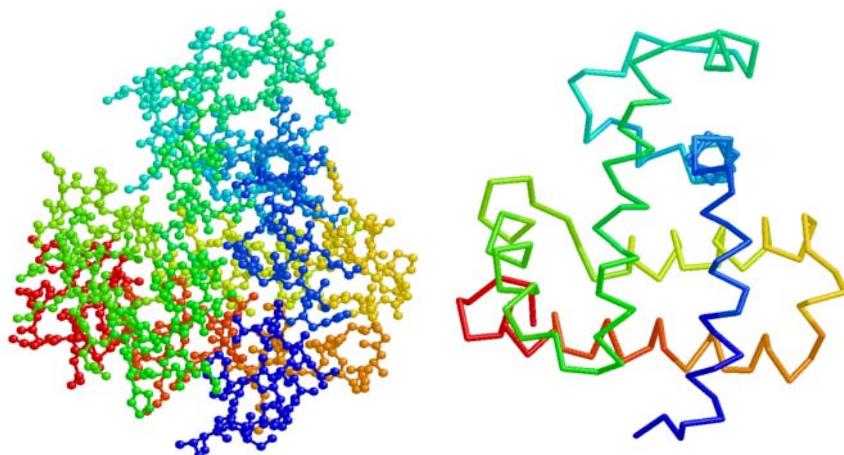
8 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

Structure alignment - Method

- What Structures Look Like?
- Structural Alignment by Iterated Dynamic Programming
 - ◊ RMS Superposition
- Scoring Structural Similarity
- Other Aspects of Structural Alignment
 - ◊ Distance Matrix based methods
 - ◊ Fold Library
- Relation of Sequence Similarity to Structural and Functional Similarity
- Protein Geometry
- Surface I (Calculation)
- Calculation of Volume
- Voronoi Volumes & Packing
- Standard Volumes & Radii
- Surfaces II (Relationship to Volumes)
- Other Applications of Volumes -- Motions, Docking

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Sperm Whale Myoglobin



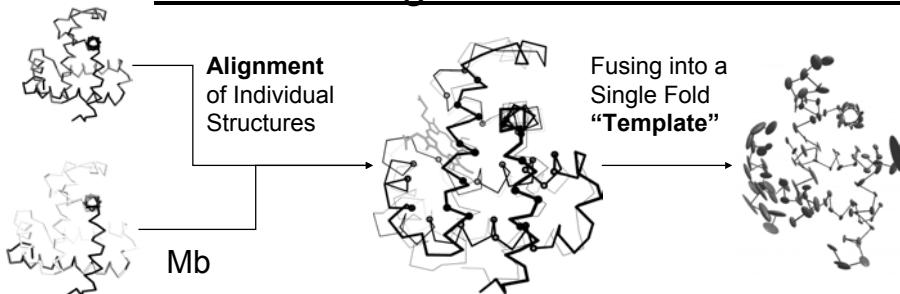
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Structural Alignment of Two Globins



11 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

Hb Automatic Alignment to Build Fold Library



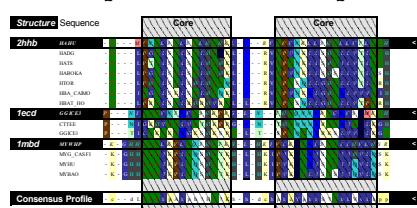
```

Hb VLSPADKTNVKAANGKVGAGAHEGYGAEEALERMFLSFPPTKTYFPH-F-DS-----HGSQAVKVGHHGKVKADALTNAV
     ||| |
Mb VLSCEGEWOLVLHVWAKVEADVGAGHGQDLIIRLFKSHPETLEKFDRFKHLKTEAMKASEDLKKHGTVTALGAIL

```

In definitions; Aligned structures, collecting together Non-homologous Sequences; Core

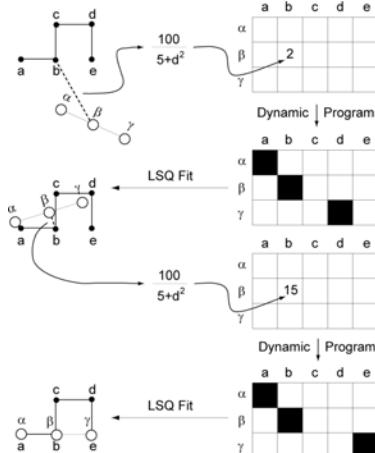
Previous work: Remington, Matthews '80; **Taylor, Orengo '89**, '94; Artymiuk, Rice, Willett '89; Sali, Blundell, '90; Vriend, Sander '91; Russell, Barton '92; **Holm, Sander '93**; Godzik, Skolnick '94; Gibrat, Madej, Bryant '96; Falicov, F Cohen, '96; Feng, Sippl '96; G Cohen '97; Singh & Brutlag, '98



Automatically Comparing Protein Structures

- Given
2 Structures (A & B),
2 Basic
Comparison Operations
 - Given an alignment optimally
SUPERIMPOSE A onto B
Find Best R & T to move A
onto B
 - Find an Alignment** between A
and B based on their 3D
coordinates

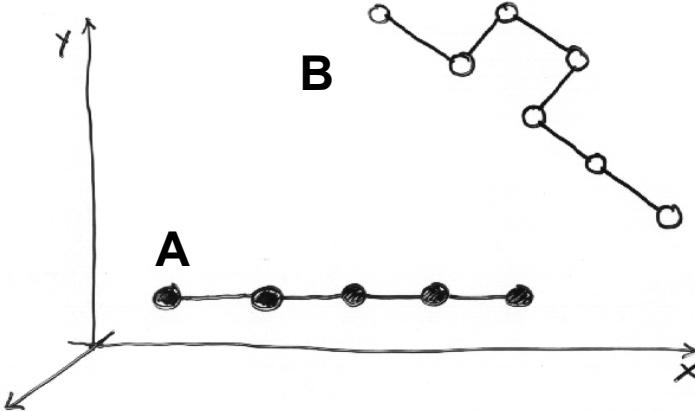
Core



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RMS Superposition (1)

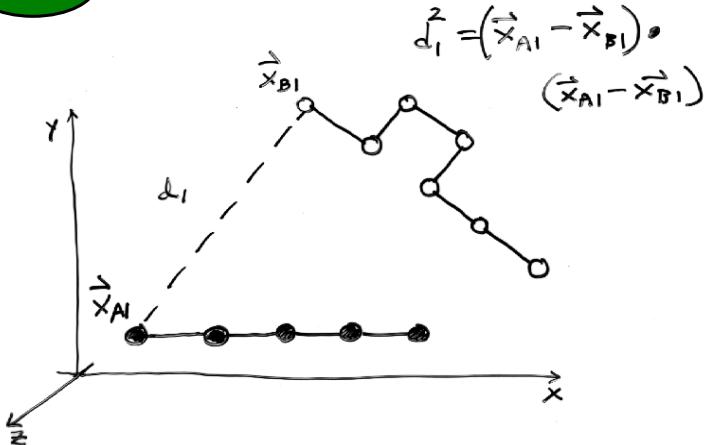
Core



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RMS Superposition (2): Distance Between an Atom in 2 Structures

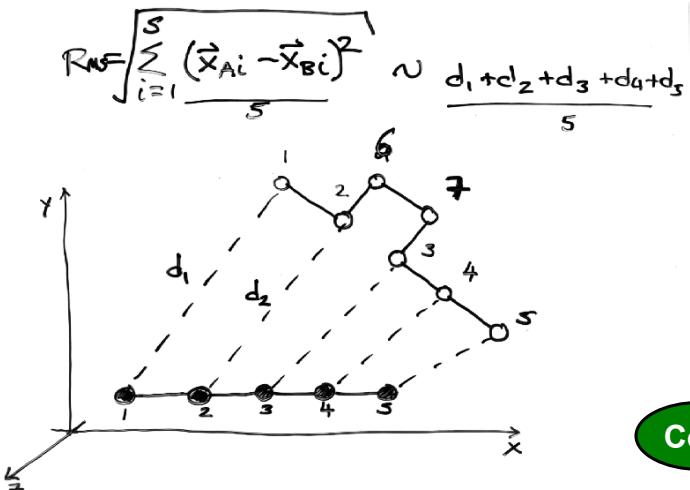
Core



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RMS Superposition (3): RMS Distance Between Aligned Atoms in 2 Structures

Core



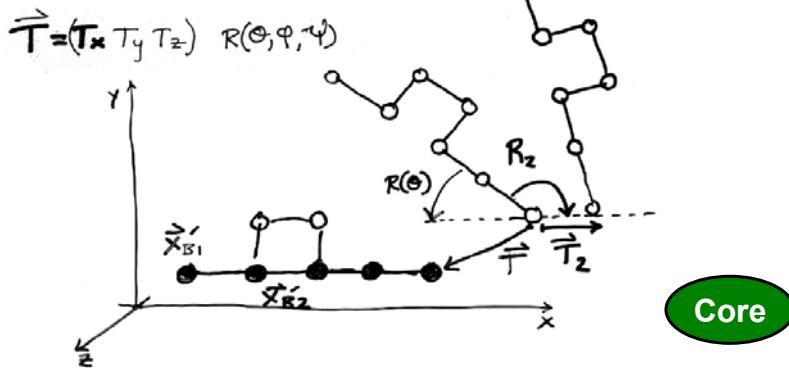
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RMS Superposition (4): Rigid-Body Rotation and Translation of One Structure (B)

$$\vec{x}'_{Bi} = R(\theta) \vec{x}_{Bi} + \vec{T}$$

ROTATE & TRANSLATE

6 parameters



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RMS Superposition (5): Optimal Movement of One Structure to Minimize the RMS

Core

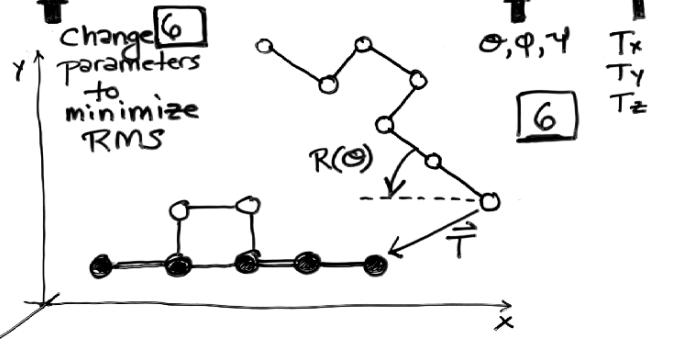
Methods of Solution:

springs
($F \sim kx$)

SVD

Kabsch

$$RMS = \sqrt{\frac{1}{N} \sum_i (\vec{x}_{Ai} - \vec{x}'_{Bi})^2} = \sqrt{\frac{1}{N} \sum_i (\vec{x}_{Ai} - R(\theta) \vec{x}_{Bi} - \vec{T})^2}$$



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Rotation Matrices

The diagram shows two coordinate systems. On the left, a 2D system rotates vector \vec{x} by angle θ around the origin to get $\vec{x}' = \begin{pmatrix} x' \\ y' \end{pmatrix}$. The rotation matrix is given as $\begin{pmatrix} \cos\theta & -\sin\theta \\ \sin\theta & \cos\theta \end{pmatrix}$. On the right, a 3D system rotates vector \vec{x} by angle θ around the z-axis to get $\vec{x}' = \begin{pmatrix} x' \\ y' \\ z' \end{pmatrix}$. The rotation matrix is given as $\begin{pmatrix} \cos\theta & -\sin\theta & 0 \\ \sin\theta & \cos\theta & 0 \\ 0 & 0 & 1 \end{pmatrix}$.

2D

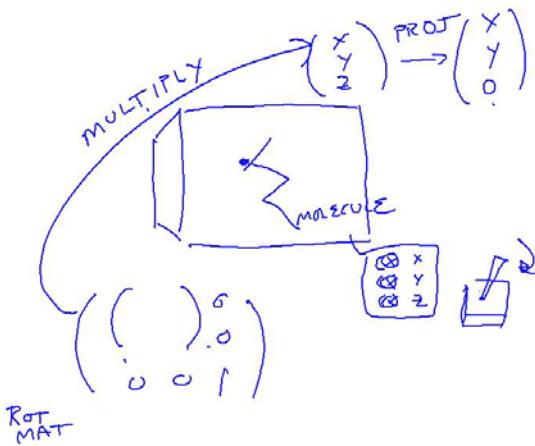
$$\begin{pmatrix} x' \\ y' \end{pmatrix} = \begin{pmatrix} \cos\theta & -\sin\theta \\ \sin\theta & \cos\theta \end{pmatrix} \begin{pmatrix} x \\ y \end{pmatrix}$$

3D

$$\begin{pmatrix} x' \\ y' \\ z' \end{pmatrix} = \begin{pmatrix} \cos\theta & -\sin\theta & 0 \\ \sin\theta & \cos\theta & 0 \\ 0 & 0 & 1 \end{pmatrix} \begin{pmatrix} x \\ y \\ z \end{pmatrix}$$

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Computer Graphics Systems: Rotation, Translation, and XY Projection



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Worked Example of Rigid Body Movement

$$\vec{x}_i = \begin{pmatrix} x_i^x \\ x_i^y \\ x_i^z \end{pmatrix} = \begin{pmatrix} 3 \\ 3 \\ 2 \end{pmatrix} = \vec{x}$$

TRANSLATE (UP BY 2)

$$\vec{x} + \vec{t} = \begin{pmatrix} 3 \\ 3 \\ 2 \end{pmatrix} + \begin{pmatrix} 0 \\ 0 \\ 2 \end{pmatrix} = \begin{pmatrix} 3 \\ 3 \\ 4 \end{pmatrix}$$

MOST ABSTRACT

$$R \vec{x}_i' = \vec{x}_i' = \begin{pmatrix} \frac{3\sqrt{3}}{2} - 3 \\ 2 \\ \frac{3}{2} + \frac{\sqrt{3}}{2} \end{pmatrix}$$

$$\begin{pmatrix} a & b & c \\ d & e & f \\ g & h & i \end{pmatrix}$$

θ, ϕ, ψ ROT BY 30° AROUND Z

PRACTICAL ABSTRACT

EX

$$\begin{pmatrix} \cos\theta & -\sin\theta & 0 \\ \sin\theta & \cos\theta & 0 \\ 0 & 0 & 1 \end{pmatrix} \begin{pmatrix} x \\ y \\ z \end{pmatrix} = \begin{pmatrix} 3 \\ 3 \\ 2 \end{pmatrix}$$

ROTATION MATRIX

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**End of class 2002.11.11
(Bioinfo-9)
[starting in sequences handout]**

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Alignment (1) Make a Similarity Matrix (Like Dot Plot)

	A	B	C	N	Y	R	Q	C	L	C	R	P	M
A	1												
Y					1								
C		1					1		1				
Y					1								
N			1										
R						1					1		
C		1					1		1				
K													
C		1					1		1				
R						1					1		
B		1											
P												1	

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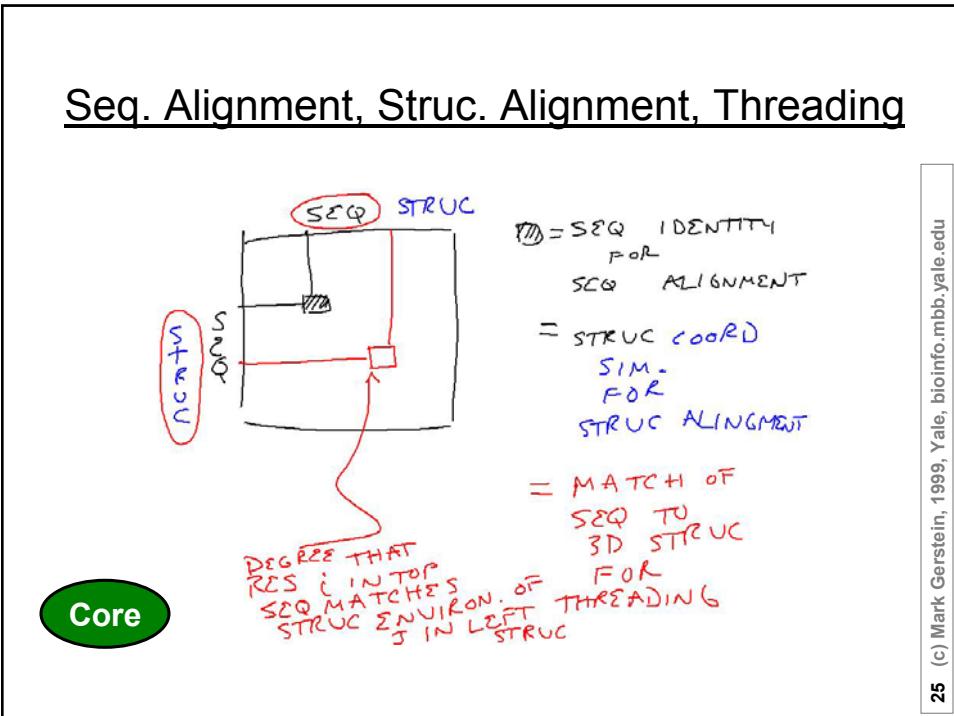
Structural Alignment (1b) Make a Similarity Matrix (Generalized Similarity Matrix)

- PAM(A,V) = 0.5
 - ◊ Applies at every position
- S(aa @ i, aa @ J)
 - ◊ Specific Matrix for each pair of residues
 - i in protein 1 and J in protein 2
 - ◊ Example is Y near N-term. matches any C-term. residue (Y at J=2)
- S(i,J)
 - ◊ Doesn't need to depend on a.a. identities at all!
 - ◊ Just need to make up a score for matching residue i in protein 1 with residue J in protein 2

	1	2	3	4	5	6	7	8	9	10	11	12	13
1	A	1											
2	Y				1			5	5	5	5	5	5
3	C		1					1		1			
4	Y				1								
5	N				1								
6	R						1					1	
7	C		1					1		1			
8	K												
9	C		1					1		1			
10	R						1					1	
11	B	1											
12	P												1

24 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu
i
J

Seq. Alignment, Struc. Alignment, Threading

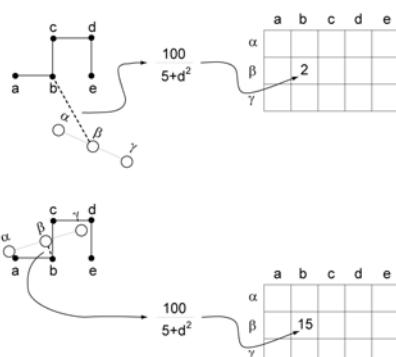


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Structural Alignment (1c*) Similarity Matrix for Structural Alignment

- Structural Alignment
 - ◊ Similarity Matrix $S(i,j)$ depends on the 3D coordinates of residues i and j
 - ◊ Distance between CA of i and j
$$d = \sqrt{(x_i - x_j)^2 + (y_i - y_j)^2 + (z_i - z_j)^2}$$

$$\diamond M(i,j) = 100 / (5 + d^2)$$
- Threading
 - ◊ $S(i,j)$ depends on the how well the amino acid at position i in protein 1 fits into the 3D structural environment at position j of protein 2



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Alignment (2): Dynamic Programming, Start Computing the Sum Matrix

```

new_value_cell(R,C) <=
    cell(R,C)                                { Old value, either 1 or 0      }
    + Max[
        cell (R+1, C+1),                      { Diagonally Down, no gaps     }
        cells(R+1, C+2 to C_max), { Down a row, making col. gap   }
        cells(R+2 to R_max, C+2) { Down a col., making row gap  }
    ]

```

	A	B	C	N	Y	R	Q	C	L	C	R	P	M
A	1												
Y					1								
C			1					1	1				
Y					1								
N				1									
R						1				1			
C			1				1	1					
K													
C			1				1	1					
R						1				1			
B		1											
P											1	0	0

	A	B	C	N	Y	R	Q	C	L	C	R	P	M
A	1												
Y						1							
C			1							1	1		
Y					1								
N				1									
R						1				1			
C			1				1	1					
K													
C			1				1	1					
R						1				2	0	0	
B	1	2	1	0	0								
P	0	1	0										

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Alignment (3):Dynamic Programming, Keep Going

	A	B	C	N	Y	R	Q	C	L	C	R	P	M
A	1												
Y					1								
C			1					1	1				
Y					1								
N				1									
R						1				1			
C			1				1	1					
K													
C			1				1	1					
R						1				2	0	0	
B	1	2	1	0	0								
P	0	1	0										

	A	B	C	N	Y	R	Q	C	L	C	R	P	M
A	1												
Y						1							
C			1							1	1		
Y						1							
N				1						1			
R						1				5	4	3	2
C	3	3	4	3	3	3	3	3	3	4	3	3	0
K	3	2	1	0									
C	2	2	3	2	2	2	2	2	3	2	3	1	0
R	2	1	1	1	1	2	1	1	1	1	2	0	0
B	1	2	1	0									
P	0												

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Alignment (4): Dynamic Programming, Sum Matrix All Done

	A	B	C	N	Y	R	Q	C	L	C	R	P	M
A	1												
Y				1									
C			1					1		1			
Y				1									
N				1									
R					5	4	3	3	3	2	2	0	0
C	3	3	4	3	3	3	3	4	3	3	1	0	0
K	3	3	3	3	3	3	3	3	3	2	1	0	0
C	2	2	3	2	2	2	2	3	2	3	1	0	0
R	2	1	1	1	1	2	1	1	1	1	2	0	0
B	1	2	1	1	1	1	1	1	1	1	1	0	0
P	0	0	0	0	0	0	0	0	0	0	0	1	0

	A	B	C	N	Y	R	Q	C	L	C	R	P	M
A	8	7	6	6	5	4	4	3	3	2	1	0	0
Y	7	7	6	6	6	4	4	3	3	2	1	0	0
C	6	6	7	6	5	4	4	4	3	3	1	0	0
Y	6	6	6	5	6	4	4	3	3	2	1	0	0
N	5	5	5	6	5	4	4	3	3	2	1	0	0
R	4	4	4	4	4	5	4	3	3	2	2	0	0
C	3	3	4	3	3	3	3	4	3	3	1	0	0
K	3	3	3	3	3	3	3	3	3	2	1	0	0
C	2	2	3	2	2	2	2	3	2	3	1	0	0
R	2	1	1	1	1	2	1	1	1	1	1	2	0
B	1	2	1	1	1	1	1	1	1	1	1	1	0
P	0	0	0	0	0	0	0	0	0	0	0	1	0

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Alignment (5): Traceback

Find Best Score (8) and Trace Back

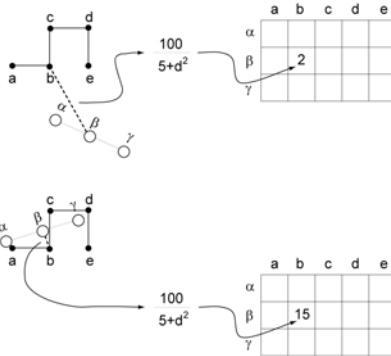
A B C N Y - R Q C L C R - P M
A Y C - Y N R - C K C R B P

	A	B	C	N	Y	R	Q	C	L	C	R	P	M
A	8	7	6	6	5	4	4	3	3	2	1	0	0
Y	7	7	6	6	6	4	4	3	3	2	1	0	0
C	6	6	7	6	5	4	4	4	3	3	1	0	0
Y	6	6	6	5	6	4	4	3	3	2	1	0	0
N	5	5	5	6	5	4	4	3	3	2	1	0	0
R	4	4	4	4	4	5	4	3	3	2	2	0	0
C	3	3	4	3	3	3	3	4	3	3	1	0	0
K	3	3	3	3	3	3	3	3	3	2	1	0	0
C	2	2	3	2	2	2	2	3	2	3	1	0	0
R	2	1	1	1	1	2	1	1	1	1	1	2	0
B	1	2	1	1	1	1	1	1	1	1	1	1	0
P	0	0	0	0	0	0	0	0	0	0	0	1	0

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In Structural Alignment, Not Yet Done (Step 6*)

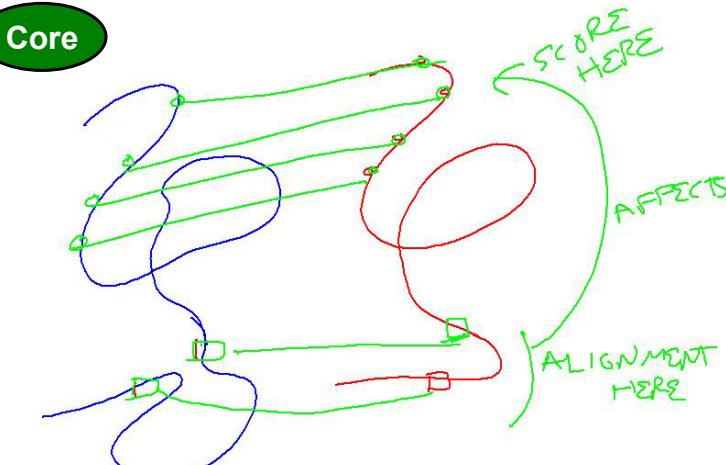
- Use Alignment to LSQ Fit Structure B onto Structure A
 - ◊ However, movement of B will now change the Similarity Matrix
- This Violates Fundamental Premise of Dynamic Programming
 - ◊ Way Residue at i is aligned can now affect previously optimal alignment of residues (from 1 to $i-1$)



ACSQRP--LRV-SH	-R	S E N C V
A-SNK PQLVKLMTH	V K	D F C V -

31 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

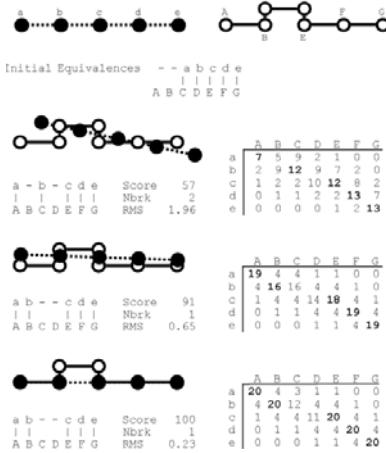
How central idea of dynamic programming is violated in structural alignment



32 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

Structural Alignment (7*), Iterate Until Convergence

- 1 Compute Sim. Matrix
- 2 Align via Dyn. Prog.
- 3 RMS Fit Based on Alignment
- 4 Move Structure B
- 5 Re-compute Sim. Matrix
- 6 If changed from #1, GOTO #2



33 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

Structure alignment - Scoring

- What Structures Look Like?
- Structural Alignment by Iterated Dynamic Programming
 - ◊ RMS Superposition
- Scoring Structural Similarity
- Other Aspects of Structural Alignment
 - ◊ Distance Matrix based methods
 - ◊ Fold Library
- Relation of Sequence Similarity to Structural and Functional Similarity
- Protein Geometry
- Surface I (Calculation)
- Calculation of Volume
- Voronoi Volumes & Packing
- Standard Volumes & Radii
- Surfaces II (Relationship to Volumes)
- Other Applications of Volumes -- Motions, Docking

34 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

Score S at End Just Like SW Score, but also have final RMS

S = Total Score

S(i,j) = similarity matrix score for aligning i and j

Sum is carried out over all aligned i and j

n = number of gaps (assuming no gap ext. penalty)

G = gap penalty

$$S = \sum_{i,j} S(i,j) - nG$$

35 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

Some Similarities are Readily Apparent others are more Subtle

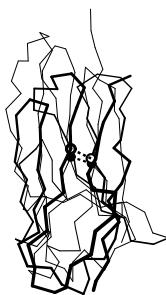
Easy:
Globins

125 res.,
~1.5 Å



Tricky:
Ig C & V

85 res.,
~3 Å

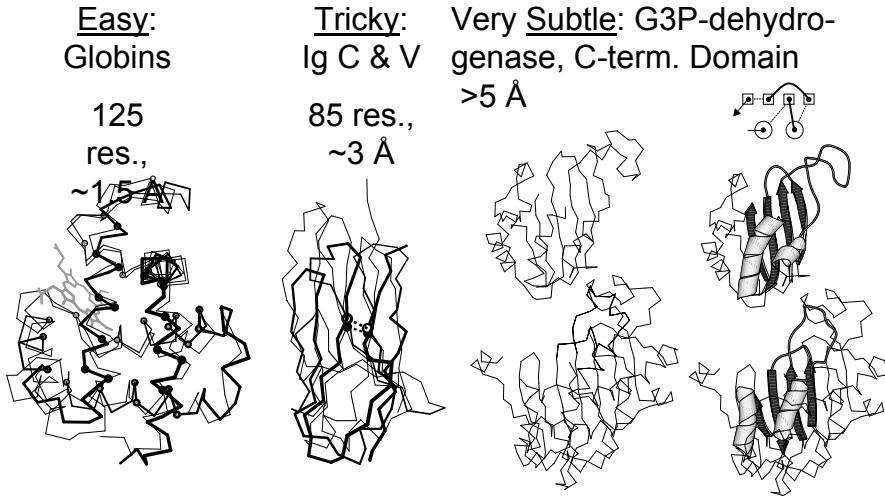


Very Subtle: G3P-dehydrogenase, C-term. Domain
>5 Å

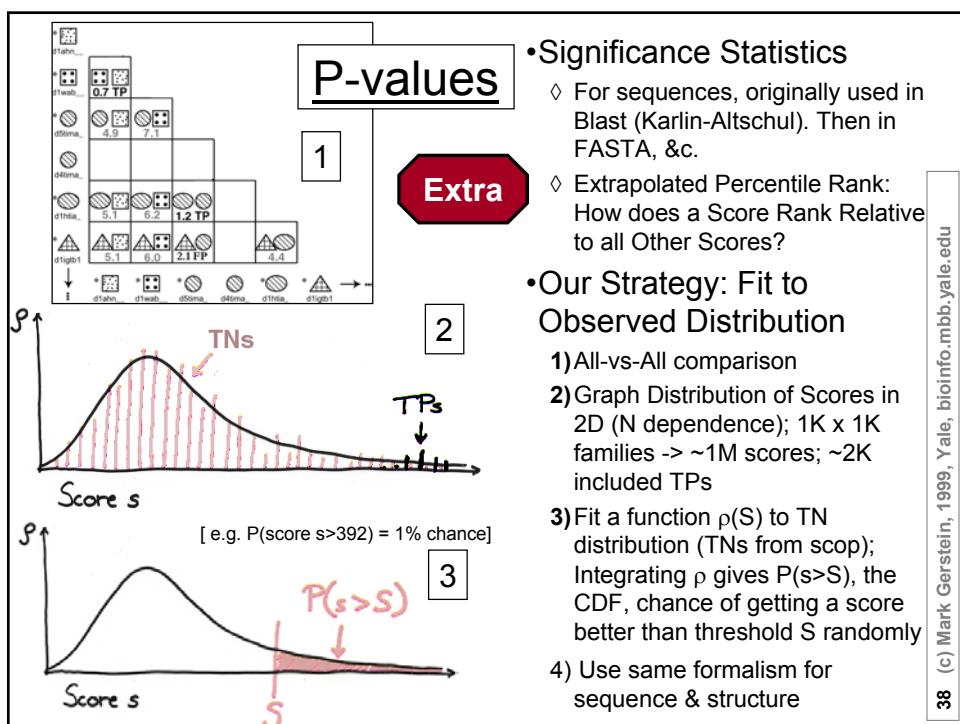


36 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

Some Similarities are Readily Apparent others are more Subtle



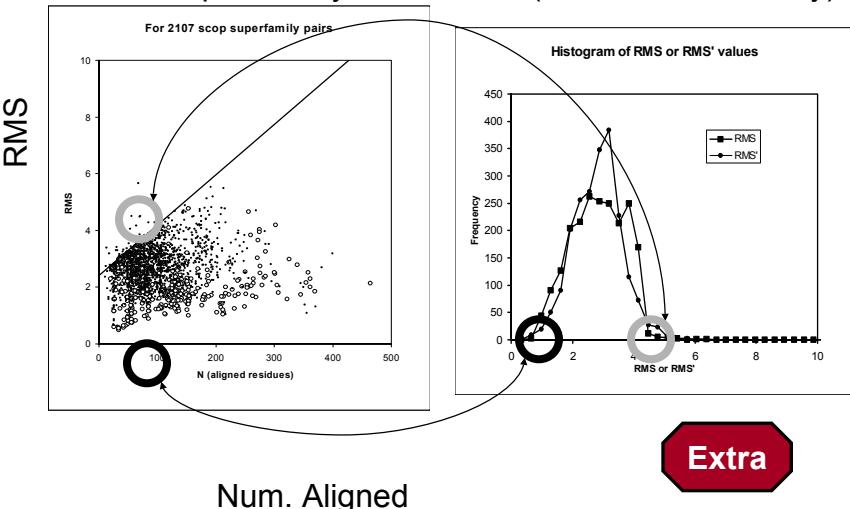
37 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu



38 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

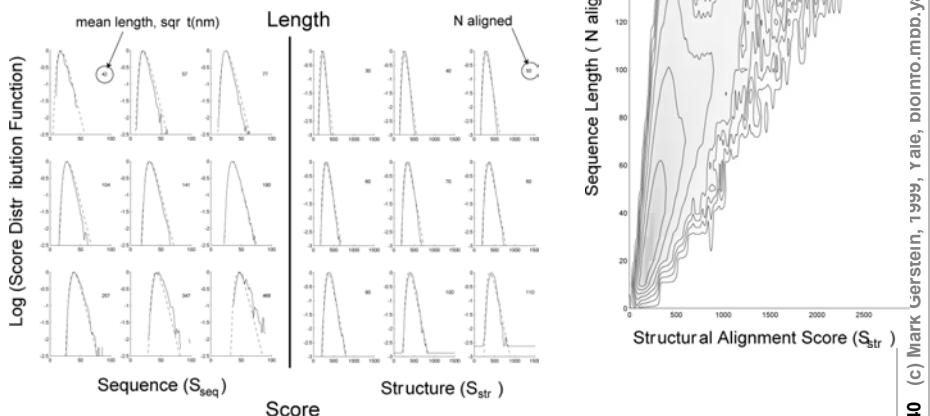
Statistics on Range of Similarities

For 2107 pairs, only 2% Outliers (with subtle similarity)



39 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

Scores from Structural Alignment Distributed Just Like Ones from Sequence Alignment (E.V.D.)



40 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

Same Results for Sequence & Structure

3 Free Parm. fit to EVD involving: a, b, σ .
These are the only difference btw. sequence and structure.

$$Z = \frac{S - (a \ln N + b)}{\sigma}$$

$$S = \sum_{i,j} M(i,j) - G$$

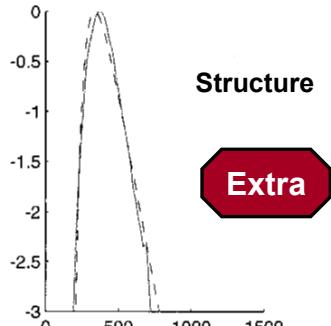
$$\rho(z) = \exp(-z - e^{-z})$$

N, G, M also defined differently for sequence and structure.

N = number of residues matched.

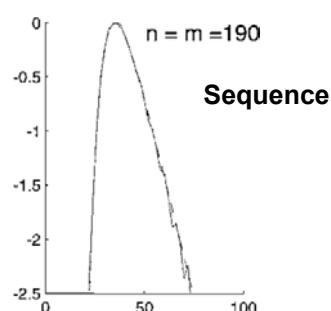
G = total gap penalty.

$M(i,j)$ = similarity matrix
(Blossum for seq. or $M_{str}(i,j)$, struc.)



Structure

Extra



Sequence

41 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

Score Significance (P-value) derived from Extreme Value Distribution (just like BLAST, FASTA)

$F(s) = E.V.D$ of scores

$$F(s) = \exp(-Z(s)) - \exp(-Z(s))$$

$$Z(s) = As + \ln(N) + B$$

s = Score from random alignment

N length of sequence matched

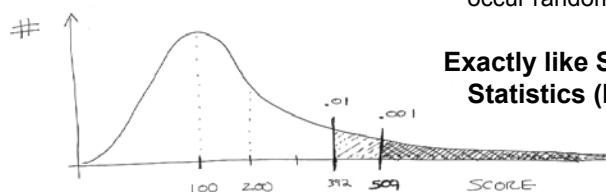
A & B are fit parameters

$$P(s > S) = CDF = \int F(s) \, ds$$

$$P(s > S) = 1 - \exp(-\exp(-Z(s)))$$

Given Score S (1%), $P(s > S)$ is the chance that a given random score s is greater than the threshold

i.e. P-value gives chance score would occur randomly



Exactly like Sequence Matching Statistics (BLAST and FASTA)

Extra

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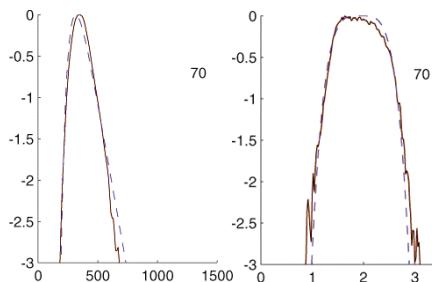
RMS is a similarity Score

Extra

- Also, RMS doesn't work instead of structural alignment (no EVD fit)
 - ◊ RMS penalizes worst fitting atoms, easily skewed

$$S_{\text{str}} \quad \text{RMS}$$

$$\sum \frac{100}{5 + d_i^2} \quad \sqrt{\sum d_i^2}$$



43 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

Structure alignment - Other methods

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Refine Method

Extra

- Multiple Alignment by aligning to central structure



- More Complex Dynamic Programming

1	i-1	i	A	B	C	D	E	F
a	x							
b		o						
c								
j-1	d							
j	e							
f							x	

AB-C-DEF
abc-de-f

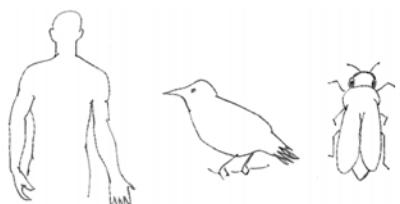
n^2 vs. n^4

- Find "best" aligned regions
 - "Core-finding" to remove outliers
 - "Noisy" suboptimal paths

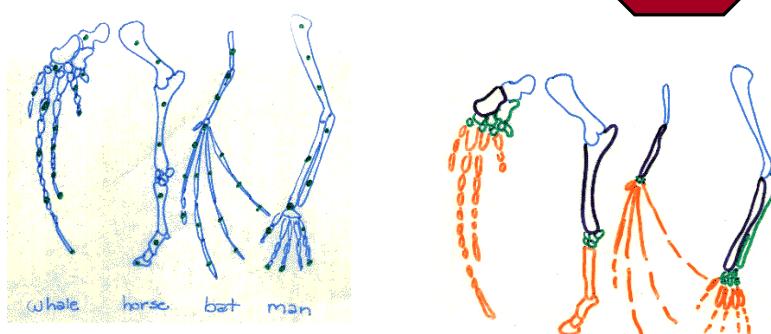
A	y	B	C	D	E
A	2	0	0	0	0
B	0	2	3	1	1
C	0	1	2	5	2
x	0	1	1	2	5
D	0	1	1	2	6
E	0	1	1	2	4

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Significance Ignoring Crucial Features in Structural Similarity



Extra



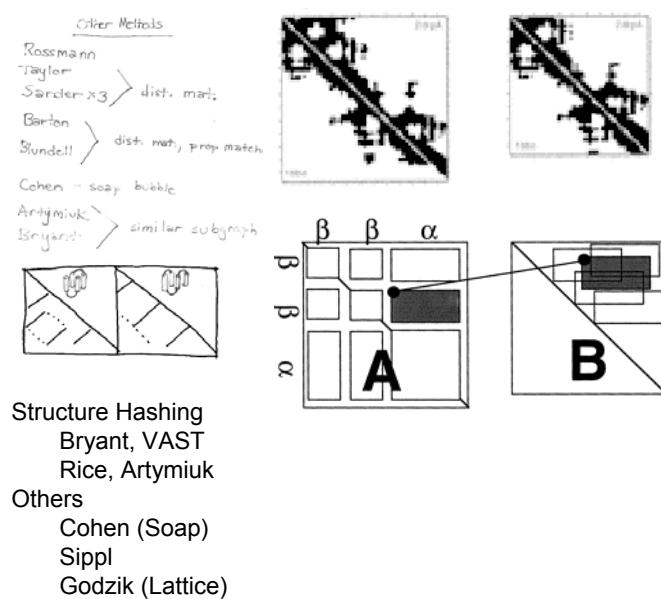
46 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

Other Methods of Structural Alignment

- RMS fitting used universally, but other alignment methods

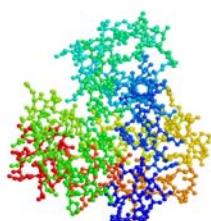
- Comparison of Distance Matrices

- ◊ Holm & Sander, DALI
- ◊ Taylor & Orengo

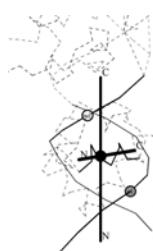


47 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

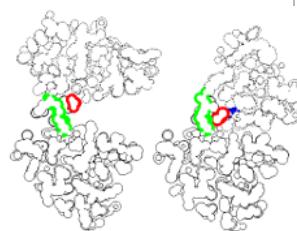
Other Aspects of Structure, Besides just Comparing Atom Positions



Atom Position,
XYZ triplets



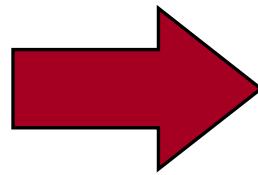
Lines, Axes,
Angles



Surfaces, Volumes

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Extra

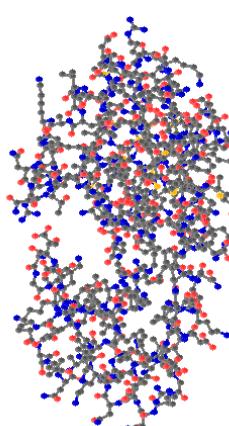


**From here to end
of Structures all is
“extra” unless
otherwise marked.**

49 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

What is Protein Geometry?

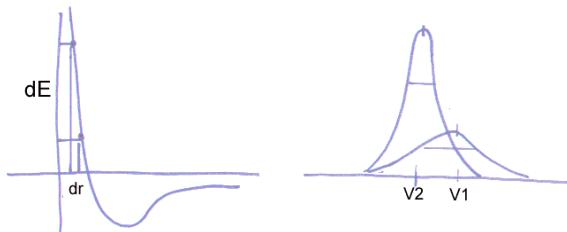
- Coordinates (X, Y, Z's)
- Derivative Concepts
 - ◊ Distance, Surface Area, Volume, Cavity, Groove, Axes, Angle, &c
- Relation to
 - ◊ Function, Energies ($E(x)$), Dynamics (dx/dt)



50 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

Small Packing Changes Significant

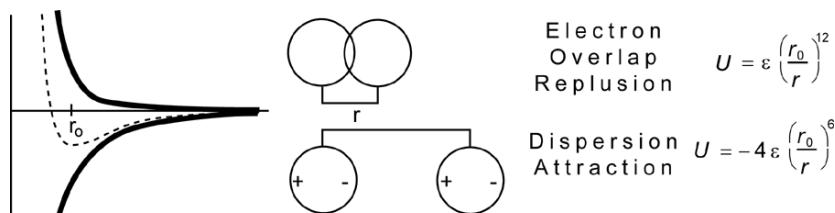
- Exponential dependence
- Bounded within a range of 0.5 (.8 and .3)
- Many observations in standard volumes gives small error about the mean (SD/sqrt(N))



51 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

Packing \sim VDW force

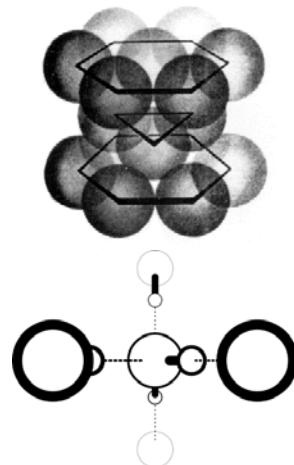
- Longer-range isotropic attractive tail provides general cohesion
- Shorter-ranged repulsion determines detailed geometry of interaction
- Billiard Ball model, WCA Theory



52 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

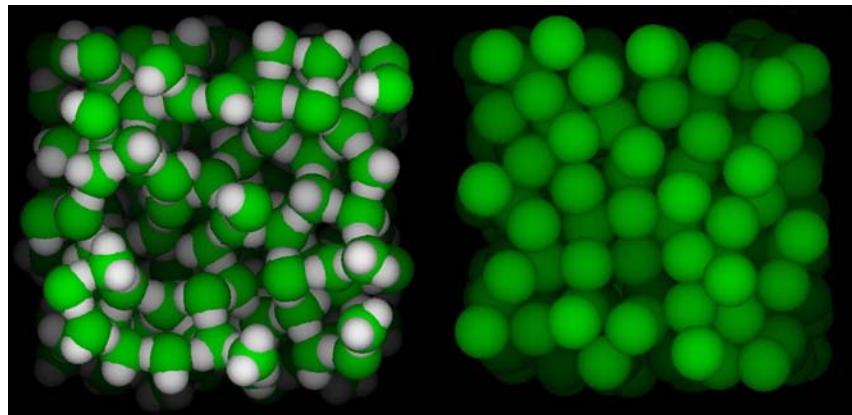
Close-packing is Default

- No tight packing when highly directional interactions (such as H-bonds) need to be satisfied
- Packing spheres (.74), hexagonal
- Water (~.35), “Open” tetrahedral, H-bonds



53 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

Water v. Argon



54 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

More Complex Systems -- what to do?

Close-Packing of Spheres

- Efficiency
 - ◊ Volume Spheres / Volume of space
- Close packed spheres
 - ◊ 74% volume filled
 - ◊ Coordination of 12
 - ◊ Two Ways of laying out
- Fcc
 - ◊ cubic close packing
 - ◊ ABC layers
- hcp
 - ◊ Hexagonally close packed
 - ◊ ABABAB

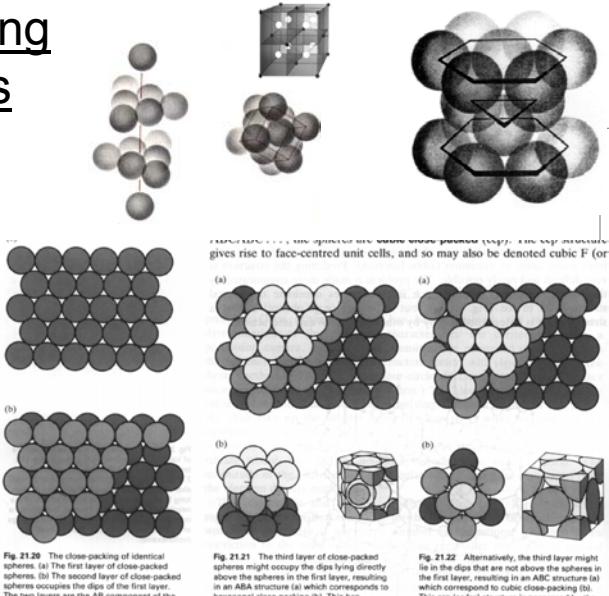
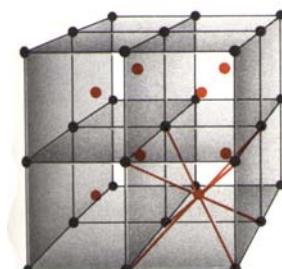
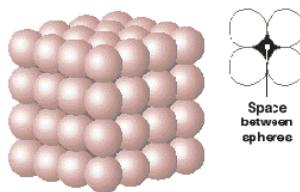


Illustration Credits: Atkins, Pchem, 634

55 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

Other Well Known Sphere Arrangements

- Simple cubic packing
 - ◊ 8 nbrs
 - ◊ 52% efficiency
- bcc cubic packing
 - ◊ one sphere sits in middle of 8 others (body-centered)
 - ◊ 8 nbrs
 - ◊ 68% efficiency
- fcc -> bcc -> simple
 - ◊ apx 3/4, 2/3, 1/2



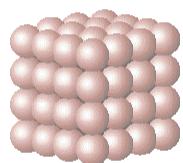
56 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

Optimal Packing Finally Proved

After Four Centuries, an Answer

What's the best way to stack a bunch of round objects? The answer, whether they are cannonballs or oranges, seems to be an extension of the familiar pyramid-shaped stock seen in grocery stores everywhere.

SIMPLE CUBIC LATTICE

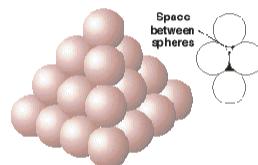


STACKING EFFICIENCY 52%

In this arrangement, the spheres sit directly on top of one another, leaving a space between the spheres that is almost equal to the sphere itself.

Stacking efficiency = volume of the spheres / (volume of the spheres + the space between the spheres)

FACE-CENTERED CUBIC LATTICE



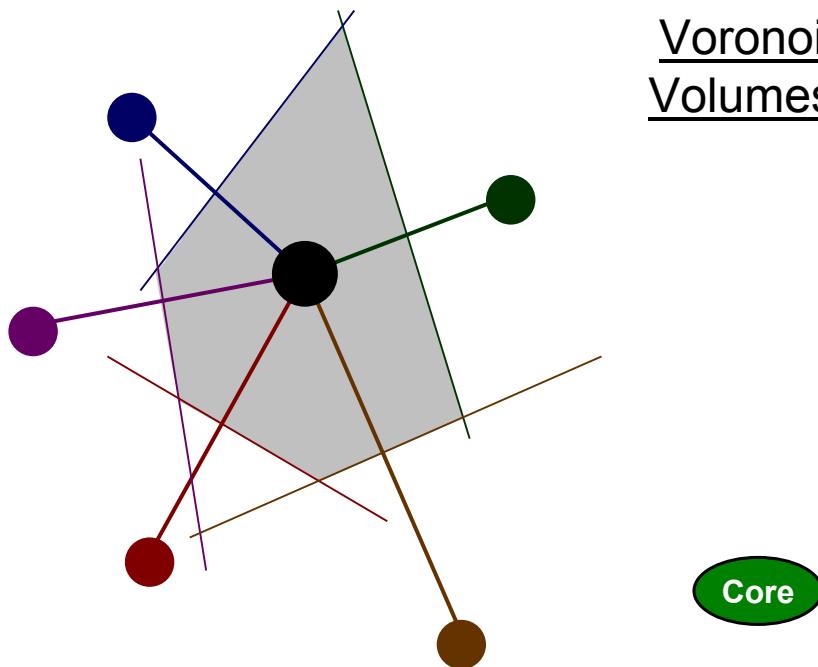
STACKING EFFICIENCY 74%

In this more efficient arrangement, the spheres sit off-center, resting within the pocket created by the spheres sitting side-by-side below.

Illustration Credits: Singh, New York Times

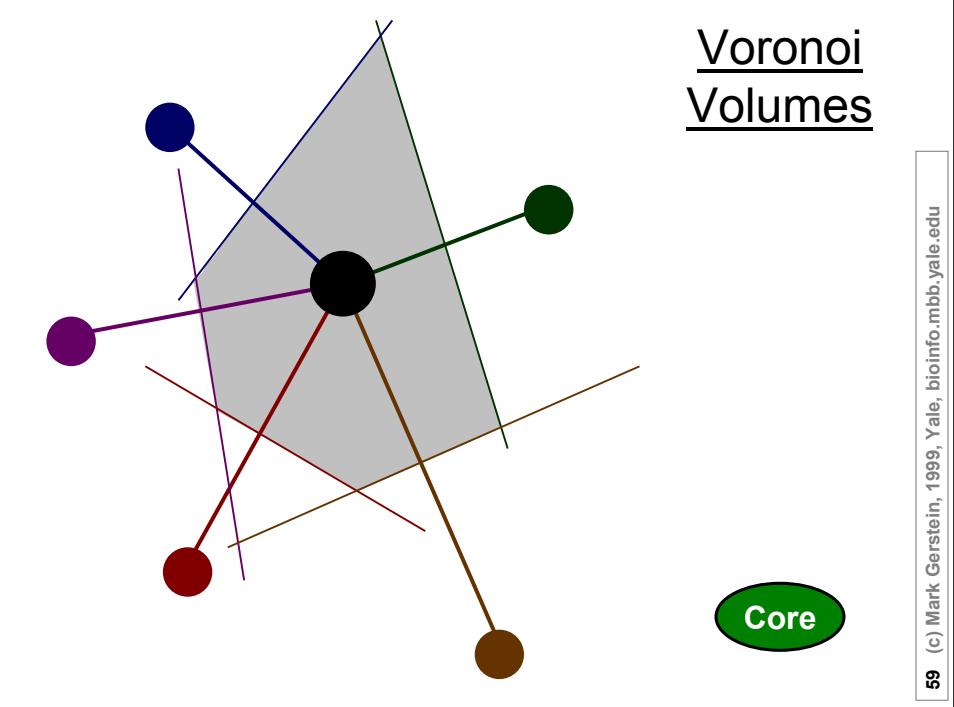
57 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

Voronoi Volumes



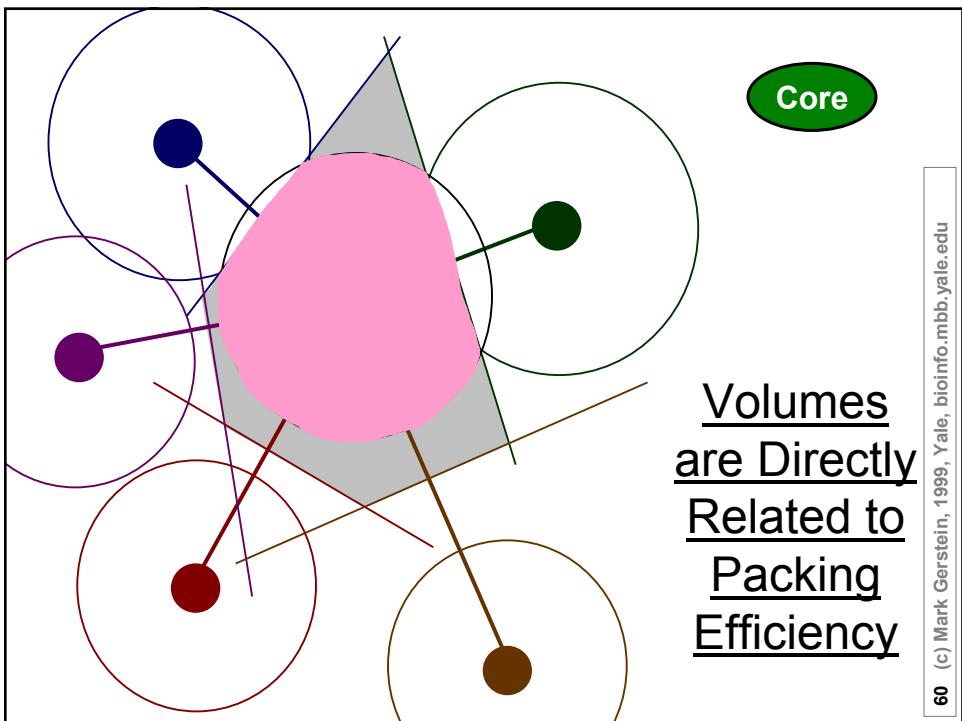
58 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

Voronoi Volumes



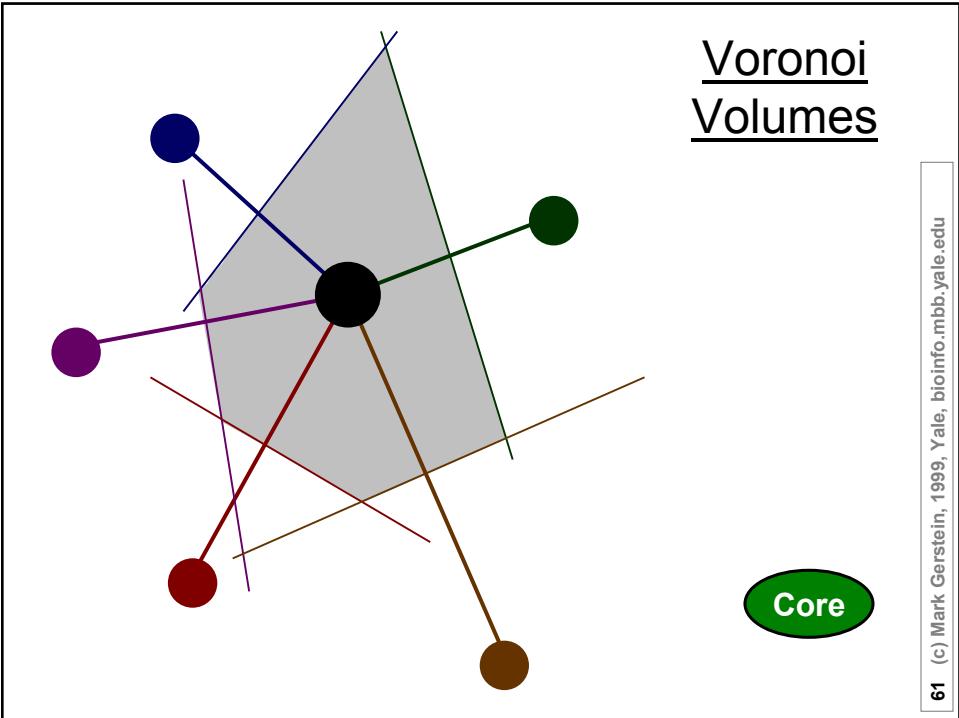
59 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

Volumes
are Directly
Related to
Packing
Efficiency



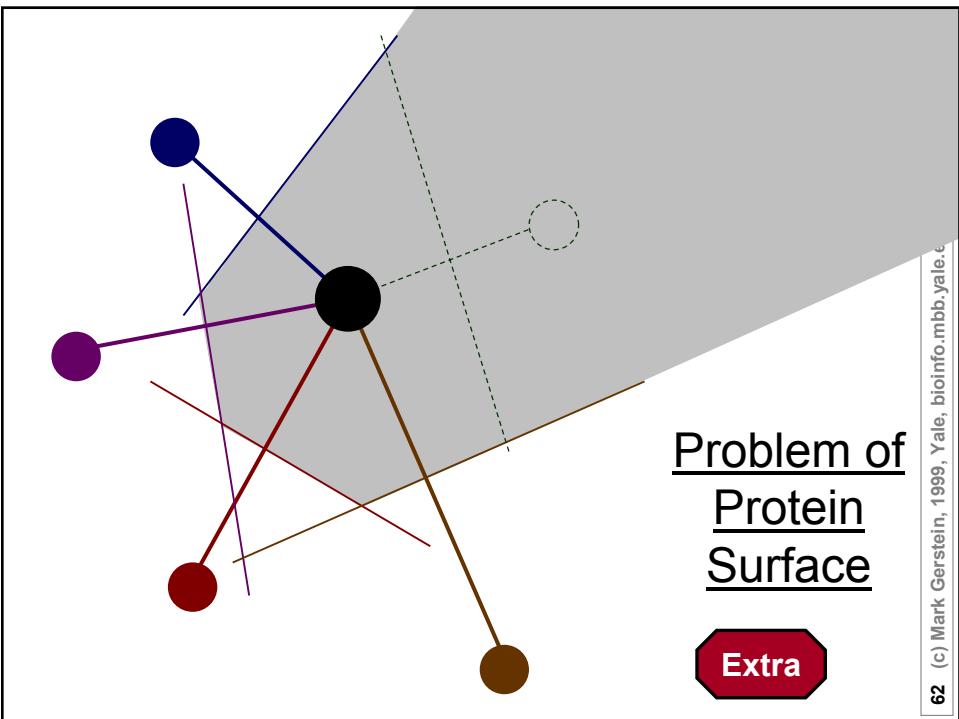
60 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

Voronoi Volumes



61 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

Problem of Protein Surface

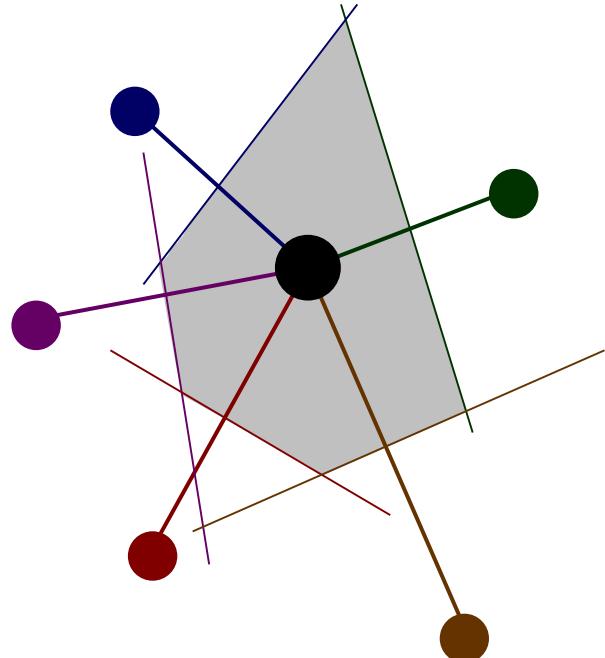


62 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

Voronoi Volumes

Extra

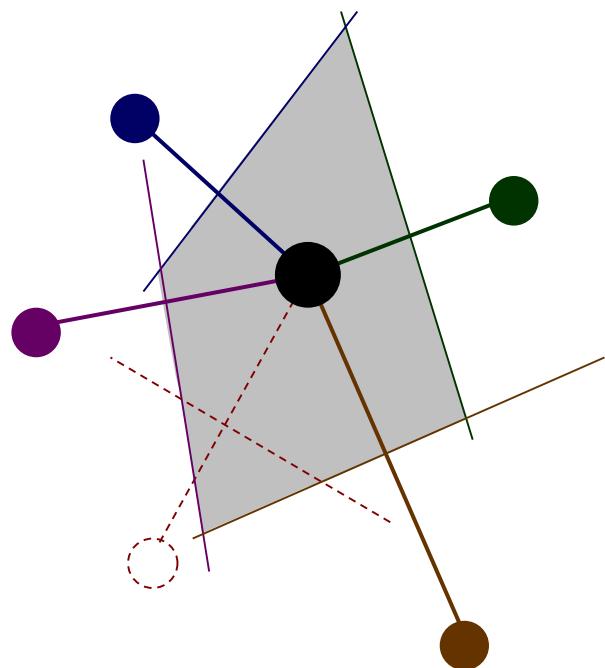
63 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu



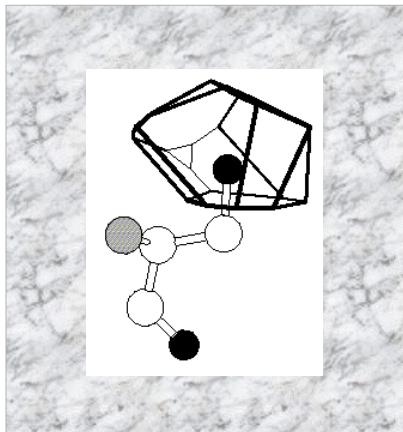
Missing Atoms Give Looser Packing

Extra

64 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu



Finding Voronoi Volumes



- Draw Lines in Between Atoms
- Draw Equidistant Planes Between Atoms
- Intersect Planes for a Volume

65 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

Classic Papers

- Lee, B. & Richards, F. M. (1971). "The Interpretation of Protein Structures: Estimation of Static Accessibility," *J. Mol. Biol.* **55**, 379-400.
- Richards, F. M. (1974). "The Interpretation of Protein Structures: Total Volume, Group Volume Distributions and Packing Density," *J. Mol. Biol.* **82**, 1-14.
- Richards, F. M. (1977). "Areas, Volumes, Packing, and Protein Structure," *Ann. Rev. Biophys. Bioeng.* **6**, 151-76.

66 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

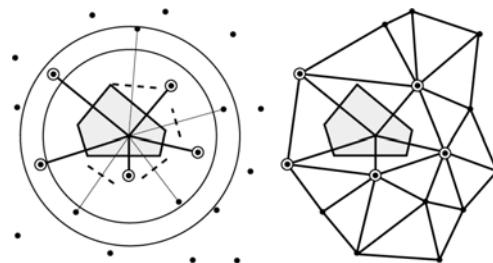
Properties of Voronoi Polyhedra

- If Voronoi polyhedra are constructed around atoms in a periodic system, such as in a crystal, all the volume in the unit cell will be apportioned to the atoms. There will be no gaps or cavities as there would be if one, for instance, simply drew spheres around the atoms.
- Voronoi volume of an atom is a weighted average of distances to all its neighbors, where the weighting factor is the contact area with the neighbor.

67 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

Delauney Triangulation, the Natural Way to Define Packing Neighbors

- Related to Voronoi polyhedra (dual)
- What “coordination number” does an atom have? Doesn’t depend on distance
- alpha shape
- threading



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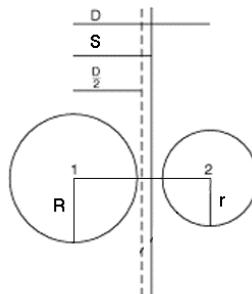
Voronoi diagrams are generally useful, beyond proteins

- Border of D.T. is Convex Hull
- D.T. produces "fatest" possible triangles which makes it convenient for things such as finite element analysis.
- Nearest neighbor problems. The nearest neighbor of a query point in center of the Voronoi diagram in which it resides
- Largest empty circle in a collection of points has center at a Voronoi vertex
- Voronoi volume of "something" often is a useful weighting factor. This fact can be used, for instance, to weight sequences in alignment to correct for over or under-representation

69 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

Atoms have different sizes

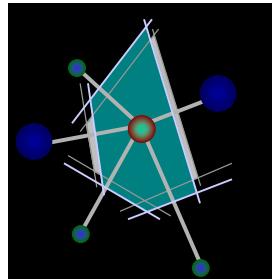
- Difficulty with Voronoi Meth.
Not all atoms created equal
- Solutions
 - ◊ Bisection -- plane midway between atoms
 - ◊ Method B (Richards)
Positions the dividing plane according to ratio
 - ◊ Radical Plane
- VDW Radii Set



70 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

Why Type the Atoms?

- Calculate Average Volumes
- Compare to Protein Atoms of Similar Type
- Allows for Modified Voronoi Volumes: Instead of Equidistant Planes, Use the Ratio of Their Radius



Courtesy of N Voss

71 (c) Mark Gerstein, 1999, Yale, bioinfo.mbb.yale.edu

Set of VDW Radii

- Great differences in a sensitive parameter (Radii for carbon 1.87 vs 2.00)
- Complex calculation: minimizing SD, iterative procedure, from protein structures
- Look for common distances in CCD
- Preliminary Solution

Atom	Bondi	New
C4	1.87	1.88
C3H1	1.76	1.76
C3H0	1.76	1.61
O1HO	1.40	1.42
O2H1	1.40	1.46
N	1.65	1.64
S	1.85	1.77

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Different Sets of Radii

Atom Type & Symbol	Bondi 1968	Lee & Richards 1971	Shrake & Rupley 1973	Richards 1974	Chothia 1975	Rich- mond & Richards 1978	Gelin & Karpplus 1979	Dunfield et al. 1979	ENCAD derived 1995	CHARMM derived 1995	Tsai et al. 1998
-CH ₃	Aliphatic, methyl	2.00	1.80	2.00	2.00	1.87	1.90	1.95	2.13	1.82	1.88
-CH-	Aliphatic, methyl	2.00	1.80	2.00	2.00	1.87	1.90	1.90	2.23	1.82	1.88
>CH-	Aliphatic, CH	-	1.70	2.00	2.00	1.87	1.90	1.85	2.38	1.82	1.88
=CH	Aromatic, CH	-	1.80	1.85	*	1.76	1.70	1.90	2.10	1.74	1.80
>C=	Trigonal, aromatic	1.74	1.80	*	1.70	1.76	1.70	1.80	1.85	1.74	1.80
-NH ₃₊	Amino, protonated	-	1.80	1.50	-	1.70	1.50	0.70	1.75	-	1.68
-NH-	Amino or amide	1.75	1.80	1.50	-	1.65	1.70	1.70	-	1.68	1.40
>NH	Peptide, NH or N	1.65	1.52	1.40	1.70	1.65	1.70	1.65	1.75	1.68	1.40
=O	Carbonyl Oxygen	1.50	1.80	1.40	1.40	1.40	1.40	1.60	1.56	1.34	1.38
-OH	Alcoholic hydroxyl	-	1.80	1.40	1.60	1.40	1.40	1.70	-	1.54	1.53
=OM	Carboxyl Oxygen	-	1.80	1.89	1.50	1.40	1.40	1.60	1.62	1.34	1.41
-SH	Sulphydryl	-	1.80	1.85	-	1.85	1.80	1.90	-	1.82	1.56
-S-	Thioether or -SS-	1.80	-	-	1.80	1.85	1.80	1.90	2.08	1.82	1.56
											1.77

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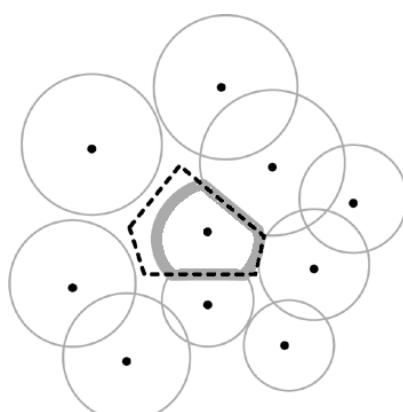
Voronoi Volumes, the Natural Way to Measure Packing

Packing Efficiency

$$= \frac{\text{Volume-of-Object}}{\text{Space-it-occupies}}$$

$$= V(\text{VDW}) / V(\text{Voronoi})$$

- Absolute v relative eff.
- V1 / V2
- Other methods
 - ◊ Measure Cavity Volume
(grids, constructions, &c)



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Standard Residue Volumes

- Database of many hi-res structures (~100, 2 Å)
- Volumes statistics for buried residues
(various selections, resample, &c)
- Standard atomic volumes harder...
parameter set development...

G 64	c 105	T 120	V 139	H 159	M 168	R 194
A 90	C 113	P 124	E 140	L 165	K 170	Y 198
S 94	D 117	N 128	N 150	I 165	F 193	W 233

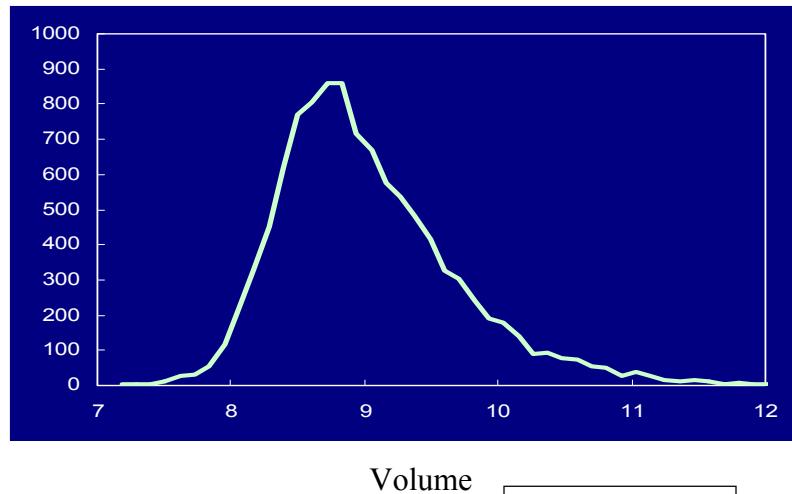
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Standard Core Volumes (Prelim.)

Atom Types	Num.	Volume (Å ³)	Error (%)
Mainchain Atoms			
carbonyl carbon (except G)	C	8361	.9.2 .08
alpha carbon (except G)	CA	7686	13.4 .09
nitrogen (except P)	N	9042	13.9 .09
carbonyl oxygen	O	7831	15.8 .10
Gly C		811	10.2 .27
Gly CA		522	23.5 .39
Pro N		334	8.6 .39
Sidechain atoms			
trigonal or aromatic carbon	>C=	3026	10.3 .13
aromatic CH (H,F,W,Y)	-CH=	4333	21.1 .14
aliphatic CH	>CH-	3411	14.6 .14
methylene group	-CH2-	5427	23.7 .12
methyl group (A,V,L,I)	-CH3	5273	36.7 .11
hydroxyl oxygen (S,T)	-OH	851	17.2 .36
carbonyl oxygen (N,Q)	=O	272	16.8 .76
carboxyl oxygen (D,E)	-O	517	16.0 .53
2° amine (R,H,W)	-NH-	530	15.6 .53
1° amine or amide (R,N,Q)	-NH2	355	23.4 .52
tetrahedral nitrogen (K)	-NH3	31	20.0 1.40
thioether or disulfide (C,M)	-S-	1242	19.3 1.22
sulphydryl (C)	-SH	67	37.8 1.33

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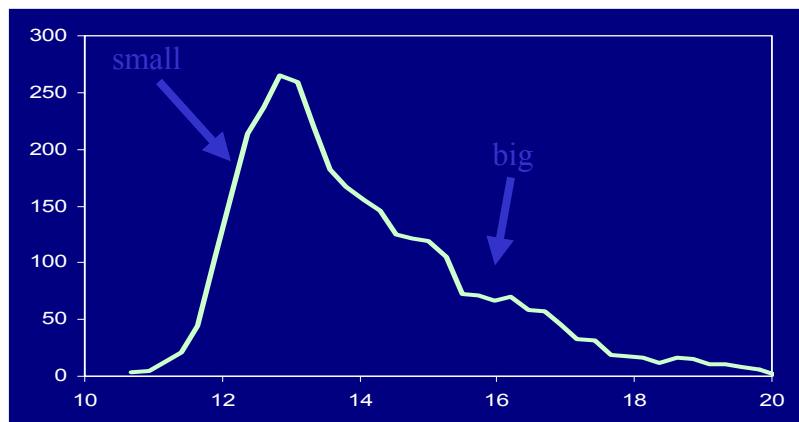
Histogram Analysis of the C₃H₀s



Courtesy of N Voss

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Histogram Analysis of the N₃H₁

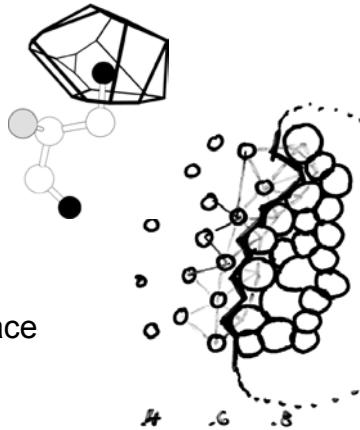


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Packing at Interfaces

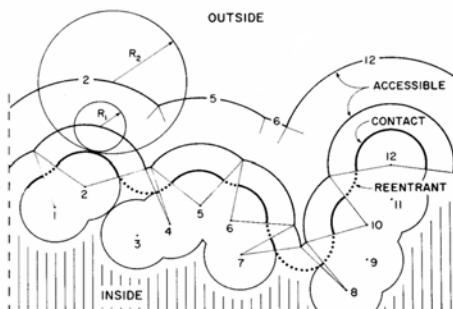
- Voronoi volumes (and D. triangulation) to measure packing
- Tight core packing v. Loose surface packing
- Grooves & ridges: close-packing v. H-bonding
- How packing defines a surface (hydration surface)
- Implications for Motions



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Richards' Molecular and Accessible Surfaces

Core



Probe Radius	Part of Probe Sphere	Type of Surface
0	Center (or Tangent)	Van der Waals Surface (vdWS)
1.4 Å	Center	Solvent Accessible Surface (SAS)
""	Tangent (1 atom)	Contact Surface (CS, from parts of atoms)
""	Tangent (2 or 3 atoms)	Reentrant Surface (RS, from parts of Probe)
""	Tangent (1,2, or 3 atoms)	Molecular Surface (MS = CS + RS)
10 Å	Center	A Ligand or Reagent Accessible Surface
∞	Tangent	Minimum limit of MS (related to convex hull)
""	Center	Undefined

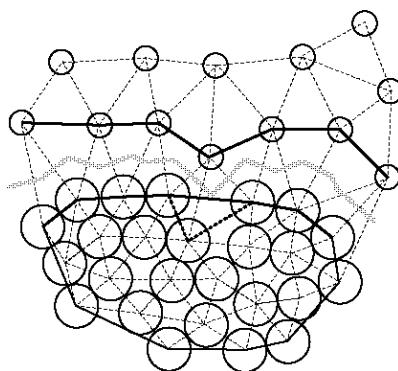
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Packing defines the “Correct Definition” of the Protein Surface

- Voronoi polyhedra are the *Natural* way to study packing!
- How reasonable is a geometric definition of the surface in light of what we know about packing
- The relationship between
 - ◊ accessible surface
 - ◊ molecular surface
 - ◊ Delauney Triangulation (Convex Hull)
 - ◊ polyhedra faces
 - ◊ hydration surface

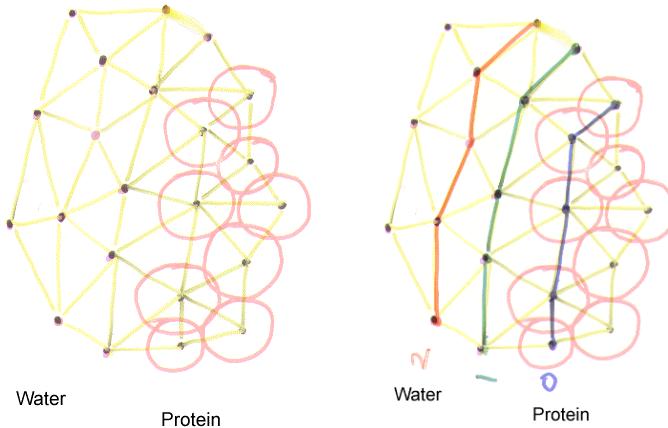
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Surface and Volume Definitions Linked



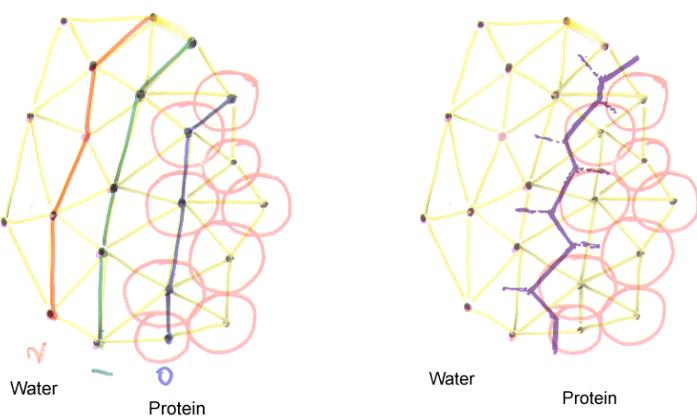
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Defining Surfaces from Packing: Convex Hull and Layers of Waters



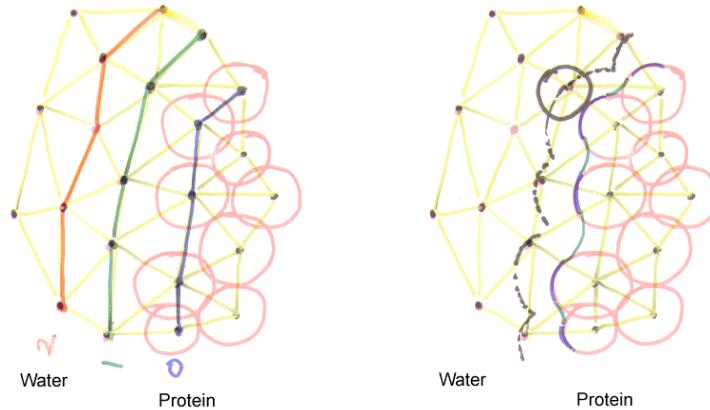
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Defining a Surface from the Faces of Voronoi Polyhedra



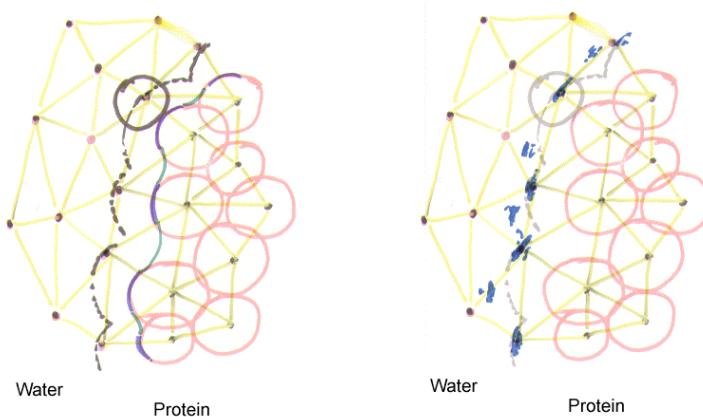
84 (c) Mark Gerstein, 1999, Yale, biolinfo.mbb.yale.edu

Accessible Surface as a Time-averaged Water Layer



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The Hydration Surface: Trying to Model Real Water



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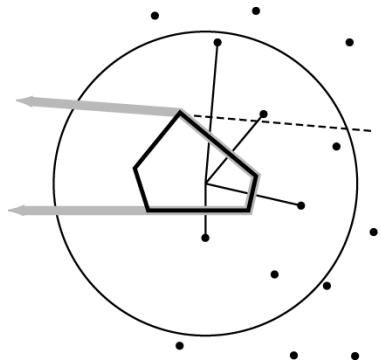
Surfaces (slides 1-10, 20-40 from website)

These are detailed slides on how to do
Voronoi construction.

Go to <http://bioinfo.mbb.yale.edu/geometry>
and follow links to “HyperTalk” tutorial on
surfaces and volumes

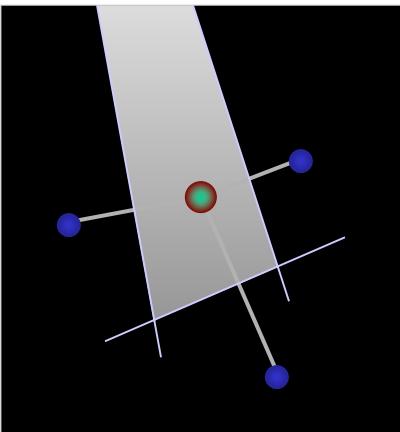
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Problem of Protein Surface for Voronoi Construction



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Problems with Voronoi Volumes



Courtesy of N Voss

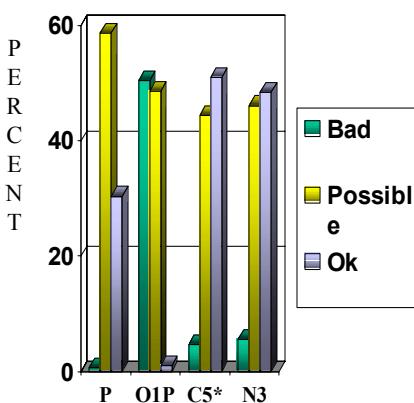
- When the Atoms are located on the Surface the Voronoi Volume fails

Ways to Correct for Unclosed Volumes

- Obtain Asymmetric Crystal Packing
- Adjust for Atom Size

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Quality of Measurement



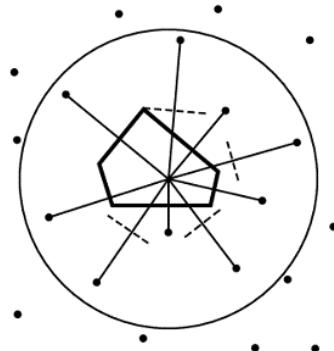
- Determine Quality By Surface Area
- If Surface Falls Below Certain Value Classified as “Ok”

Courtesy of N Voss

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Voronoi Volumes

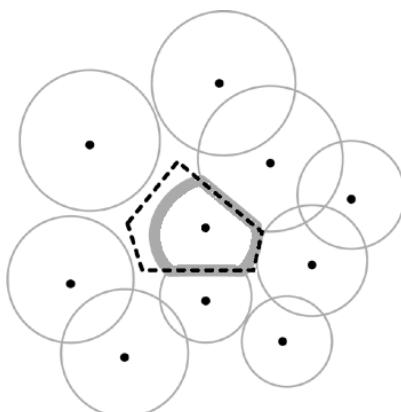
- Each atom surrounded by a single convex polyhedron and allocated space within it
 - ◊ Allocation of all space (large V implies cavities)
- 2 methods of determination
 - ◊ Find planes separating atoms, intersection of these is polyhedron
 - ◊ Locate vertices, which are equidistant from 4 atoms



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Voronoi Volumes, the Natural Way to Measure Packing

- Packing Efficiency
- $$\begin{aligned} &= \frac{\text{Volume-of-Object}}{\text{Space-it-occupies}} \\ &= \frac{V(\text{VDW})}{V(\text{Voronoi})} \\ \bullet &\text{ Absolute v relative eff.} \\ &V_1 / V_2 \\ \bullet &\text{ Other methods} \\ &\diamond \text{ Measure Cavity Volume} \\ &\quad (\text{grids, constructions, \&c}) \end{aligned}$$



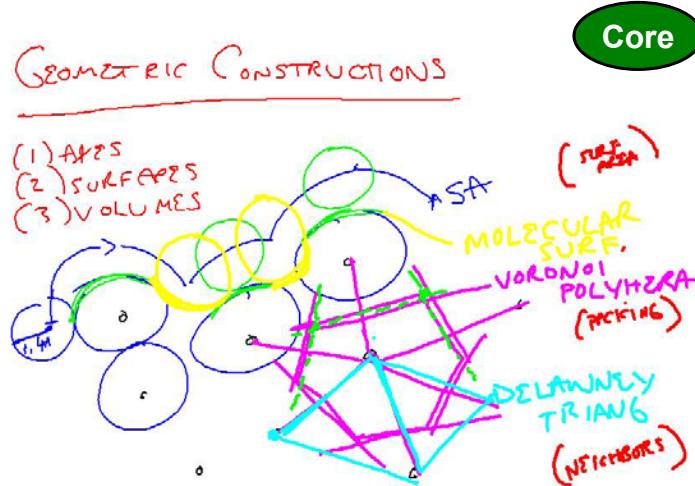
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Calculating Volumes with Voronoi polyhedra

- In 1908 Voronoi found a way of partitioning all space amongst a collection of points using specially constructed polyhedra. Here we refer to a collection of "atom centers" rather than "points."
- In 3D, each atom is surrounded by a unique limiting polyhedron such that all points within an atom's polyhedron are closer to this atom than all other atoms.
- Likewise, points equidistant from 2 atoms form planes (lines in 2D). Those equidistant from 3 atoms form lines, and those equidistant from 4 centers form vertices.

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Summary of Geometric Constructions



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**End of class 2002,11.13
(Bioinfo-10)
[Next is discussion of
expression]**